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(71) Applicant (for all designated States except US): CONNAUGHT LABORATORIES LIMITED [CA/CA]; 1755 Steeles Avenue West, Willowdale, Ontario M2R 3T4 (CA).			
(72) Inventors; and			
(75) Inventors/Applicants (for US only): CHONG, Pele [CA/CA]; 32 Estoril Street, Richmond Hill, Ontario L4C 0B6 (CA). THOMAS, Wayne [AU/AU]; 31 Taylor Road, Nedlands, W.A. 6009 (AU). YANG, Yan-Ping [CN/CA]; Apt. 1709, 120 Torresdale Avenue, Willowdale, Ontario M2R 3N7 (CA). LOOSMORE, Sheena [CA/CA]; 70 Crawford Rose Drive, Aurora, Ontario L4G 4R4 (CA). SIA, Dwo, Yuan, Charles [CA/CA]; 189 Mabley Crescent, Thornhill, Ontario L4J 2Z7 (CA). KLEIN, Michel [CA/CA]; 16 Munro Boulevard, Willowdale, Ontario M2P 1B9 (CA).			
(74) Agent: STEWART, Michael, I.; Sim & McBurney, 330 University Avenue, Suite 701, Toronto, Ontario M5G 1R7 (CA).			

(54) Title: HAEMOPHILUS OUTER MEMBRANE PROTEIN

(57) Abstract

Purified and isolated nucleic acid from specific strains of *Haemophilus influenzae* is provided which encodes at least a portion of the D15 outer membrane protein of *Haemophilus*. The nucleic acid is used to produce peptides, polypeptides and proteins free of contaminant associated with *Haemophilus* for purposes of diagnosis and medical treatment. Furthermore, the nucleic acid may be used in the diagnosis of *Haemophilus* infection. Antisera obtained following immunization with the nucleic acid D15 outer membrane protein or peptides also may be used for the purpose of diagnosis and medical treatment.

D15 SEQUENCE COMPARISON

.....	Ca
.....	Eagen
.....	Murra
.....	SB33
.....	PAK
.....
.....	Ca
.....	Eagen
.....	Murra
.....	SB33
.....	PAK
.....
.....	Ca
.....	Eagen
.....	Murra
.....	SB33
.....	PAK
.....
.....	Ca
.....	Eagen
.....	Murra
.....	SB33
.....	PAK
.....
.....	Ca
.....	Eagen
.....	Murra
.....	SB33
.....	PAK
.....
.....	Ca
.....	Eagen
.....	Murra
.....	SB33
.....	PAK

TITLE OF INVENTIONHAEMOPHILUS OUTER MEMBRANE PROTEINFIELD OF INVENTION

5 The present invention is related to the field of molecular genetics and is particularly concerned with the cloning of an outer membrane protein D15 of Haemophilus.

BACKGROUND OF THE INVENTION

10 Haemophilus influenzae type b (Hib) is a major cause of bacterial meningitis in children under the age of five years. Protective antibodies to the disease are induced by the capsular polysaccharide of the organism and a vaccine was developed that utilises the purified polyribosyl ribitol phosphate (PRP) as the antigen. This
15 vaccine provides 90% protection in adults and in children over 24 months of age, but was ineffective in children under 24 months Zangwill et al 1993 (The references are identified in a list of reference at the end of this disclosure). Like other polysaccharide antigens, PRP
20 does not induce the proliferation of T-helper cells, and re-immunisation fails to elicit either a booster response or an increase in memory cells. Conjugation of the PRP polysaccharide with protein carriers confers T-cell dependent characteristics to the vaccine and
25 substantially enhances the immunologic response to the PRP antigen. Currently, there are four PRP-carrier conjugate vaccines available. These are vaccines based upon H. influenzae type b capsular polysaccharide conjugated to diphtheria toxoid, tetanus toxoid, or
30 Neisseria meningitidis outer membrane protein (reviewed in Zangwill et al 1993).

 However, the current Haemophilus conjugate vaccines only protect against meningitis caused by Haemophilus influenzae type b. They do not protect against other
35 invasive typeable strains (types a and c) and, more importantly, against non-typeable (NTHi) strains which are a common cause of postpartum and neonatal sepsis,

otitis media, epiglottitis, pneumonia, and tracheobronchitis, are required.

SUMMARY OF THE INVENTION

The present invention is directed towards the provision of purified and isolated nucleic acid molecules comprising at least a portion coding for a D15 outer membrane protein of a species of Haemophilus. The nucleic acid molecules comprising at least a portion coding for D15 outer membrane protein are useful for the specific detection of strains of Haemophilus, and for diagnosis of infection by Haemophilus. The purified and isolated nucleic acid molecules, such as DNA comprising at least a portion coding for D15 outer membrane protein, are also useful for expression of the D15 gene by recombinant DNA means for providing, in an economical manner, purified and isolated D15 outer membrane protein.

The D15 outer membrane protein or fragments thereof or analogs thereof are useful immunogenic compositions for the preparation of vaccines against diseases caused by Haemophilus, the diagnosis of infection by Haemophilus and as tools for the generation of immunological reagents. Mono- or polyclonal antisera (antibodies) raised against the D15 outer membrane protein produced in accordance with aspects of the present invention are useful for the diagnosis of infection by Haemophilus, specific detection of Haemophilus (in, for example, in vitro and in vivo assays) and for the treatment of diseases caused by infection by Haemophilus.

Peptides corresponding to portions of the D15 outer membrane protein or analogs thereof are useful immunogenic compositions for the preparation of vaccines against disease caused by Haemophilus, the diagnosis of infection by Haemophilus and as tools for the generation of immunological reagents. Mono- or polyclonal antisera raised against these peptides, produced in accordance with aspects of the present invention, are useful for the

comprises at least an 18 bp fragment selected from the DNA molecules as recited above is inserted. The recombinant plasmid may be plasmid DS-712-2-1 having ATCC accession number 75604, deposited November 4, 1993 and
5 plasmid JB-1042-5-1 having ATCC accession number 75006, deposited November 4, 1993.

The plasmids may be adapted for expression of the encoded D15 outer membrane protein in a host cell, which may be a heterologous or homologous host, by
10 incorporation into a recombinant vector, provided in accordance with a further aspect of the invention. The recombinant vector may comprise at least a DNA segment comprising at least an 18 bp fragment selected from the DNA molecules as recited above and expression means
15 operatively coupled to the DNA segment for expression of the gene product encoded thereby in the host cell. The plasmid for expression of the encoded D15 outer membrane protein may be plasmid DS-880-1-2 having ATCC accession number 75605, deposited November 4, 1993 being adapted
20 for expression at the D15 outer membrane protein in E. coli. The selected DNA segment may encode a polypeptide of at least 6 residues and, in particular, may be selected from those segments encoding a polypeptide of Table 2 (below). The DNA segment may further comprise a
25 nucleic acid sequence encoding a leader sequence for export of the gene product from the host. The host for expression may be selected from, for example, Escherichia coli, Bacillus, Haemophilus, fungi, yeast or the baculovirus expression system may be used.

30 Additional aspects of the invention include the protein encoded by the DNA molecule comprising at least a portion coding for the D15 outer membrane protein, fragment or a functional analog of such protein, the use of the protein or analog in vaccination and diagnosis,
35 and the generation of immunological reagents. The invention also includes antisera (antibodies) raised

of the immunogenic composition or the nucleic acid molecule as recited above to provide protective immunity against Haemophilus infection.

5 The present invention further includes a chimeric molecule comprising a D15 protein or peptide corresponding thereto as provided herein linked to another polypeptide or protein or a polysaccharide. The linked polypeptide or protein may comprise a surface protein or peptide corresponding thereto from a pathogenic bacteria, which may be the P1, P2 or P6 outer
10 membrane protein of H. influenzae. The linked polysaccharide preferably comprise a PRP molecule from H. influenzae.

BRIEF DESCRIPTION OF THE FIGURES

15 The present invention will be further understood from the following description with reference to the drawings, in which:

Figure 1A shows the nucleotide sequence of the D15 gene from H. influenzae type b Ca strain (SEQ ID NO: 1) and its deduced amino acid sequence (SEQ ID NO: 2);
20

Figure 1B shows the nucleotide sequence of the D15 gene from H. influenzae type b Eagan strain (SEQ ID NO. 3) and its deduced amino acid sequence (SEQ ID NO: 4);

Figure 1C shows the nucleotide sequence of the D15 gene from H. influenzae type b MinnA strain (SEQ ID NO. 5) and its deduced amino acid sequence (SEQ ID NO: 6);
25

Figure 1D shows the nucleotide sequence of the D15 gene from H. influenzae non-typeable SB33 (SEQ ID NO. 7) and its deduced amino acid sequence (SEQ ID NO: 8);

Figure 1E shows the nucleotide sequence of the D15 gene from H. influenzae non-typeable PAK 12085 (SEQ ID NO. 9) and its deduced amino acid sequence (SEQ ID NO: 10);
30

Figure 1F shows an alignment of the nucleotide sequences of the D15 genes (SEQ ID NOS: 1, 3, 5, 7 and 9)
35

97kDa); 2, GST standard; 3, GST-(D15 fragment) fusion protein; 4, fusion protein cleaved by thrombin; 5, N-terminal rD15 fragment; 6, GST; 7, low molecular weight markers;

5 Figure 10 shows guinea pig IgG antibody response to N-terminal rD15 fragment. The arrows indicate the immunization schedule. Bleeds were taken at 2, 4, 6 and 8 weeks. The bars represent the standard deviation; and

10 Figure 11 shows the hydrophilicity plot of D15 established by using a window average across 7 residues according to Hope, 1986.

GENERAL DESCRIPTION OF THE INVENTION

Any Haemophilus strains that have D15 genes may be conveniently used to provide the purified and isolated
15 nucleic acid molecules (which may be in the form of DNA molecules), comprising at least a portion coding for a D15 outer membrane protein as typified by embodiments of the present invention. Such strains are generally available from clinical sources and from bacterial
20 culture collections, such as the American Type Culture Collection. H. influenzae strains may include types a, b and c strains, non-typeable strains and other bacteria that produce a D15 protein, fragment or analog thereof. Appropriate strains of Haemophilus include:-

25 H. influenzae type b strain Ca;
 H. influenzae type b strain MinnA;
 H. influenzae type b strain Egan;
 H. influenzae non-typeable b strain SB33; or
 H. influenzae non-typeable b strain PAK 12085.

30 In this application, the term D15 outer membrane protein is used to define a family of D15 proteins which includes those having naturally occurring variations in their amino acid sequences as found in various strains of, for example, Haemophilus. The purified and isolated
35 DNA molecules comprising at least a portion coding for D15 outer membrane protein of the present invention also

described herein are advantageous as diagnostic reagents, antigens for the production of Haemophilus-specific antisera, for vaccination against the diseases caused by species of Haemophilus and for detecting infection by Haemophilus.

Reference will now be made in detail to the presently preferred embodiments of the invention, which together with the following Examples, serve to explain the principle of the invention. For clarity of disclosure, and not by way of limitation, the detailed description of the invention is divided into the following sections:

(i) The DNA sequences coding for the outer membrane protein D15 from H. influenzae type b Ca strain.

A clone producing the outer membrane protein designated D15 of H. influenzae type b (Hib) was isolated by screening a genomic library with H. influenzae type b OMP-specific polyclonal antibodies as previously described by Berns and Thomas 1965; Thomas and Rossi 1986. The DNA fragment encoding the D15 protein was isolated, subcloned into pUC19 to produce pUC19/D15 (Figure 2) and used to transform E. coli HB101 as described in Example 1. Plasmid DNA was prepared from two individual colonies of E. coli HB101 containing the pUC19/D15 plasmid. Sequencing was performed on an ABI DNA sequencer model 370A using dye-terminator chemistry and oligonucleotide primers which had been synthesized on an ABI DNA synthesizer model 380B, and purified by chromatography. Nucleotide sequence analysis of the D15 gene revealed that it contains a putative promoter and an open reading frame encoding 789 amino acids (Figure 1A).

The first 19 amino acid residues of the translated open reading frame form a typical leader sequence as found in other H. influenzae type b outer membrane proteins, such as P1 and P2. The N-terminal sequence of immuno-affinity purified native D15 antigen was

heterologous proteins in E. coli. The T7 expression system is described in U.S. Patent 4,952,496. Clones were, therefore, constructed which utilize the T7 system to express a mature D15 protein that contains an additional methionine residue at the amino terminus. The D15 signal sequence was removed during this construction process. A full length recombinant D15 (termed rD15) was expressed in inclusion bodies which allow the D15 protein to be readily purified. The D15 genes from H. influenzae type b strain Ca and H. influenzae non-typeable SB33 strain have been expressed at high levels in E. coli using the T7 system to permit production of large quantities of rD15 protein. The construction of clone DS-880-1-2 which expresses the SB33 D15 gene is described herein (see Figure 4 and Example 5). The rD15 protein was immunologically similar to its native counterpart isolated from H. influenzae typeable and non-typeable strains (see below). Thus, rD15 may be used as a cross-reactive antigen in a diagnostic kit to detect many, if not all, strains of H. influenzae and other bacteria that produce a D15 outer membrane protein or analog thereof. Alternatively, rD15 can be used as an antigen to specifically detect the presence of H. influenzae in a sample.

25 A truncated D15 fragment was expressed in E. coli as a fusion protein with glutathione S-transferase (GST), as described in Example 6. The construction was designed to express the N-terminal fragment of the D15 protein. The fusion protein was expressed at high levels from a pGEX-2T construction and the N-terminal fragment was cleaved from the GST carrier protein by treatment with thrombin. This procedure generated a molecule termed the N-terminal rD15 fragment which encompasses amino acids 63-223 of the D15 protein. This N-terminal rD15 fragment was highly immunogenic and elicited protective antibodies against challenge with live H. influenzae.

urea (see Example 8). After dialysis against PBS to remove urea, more than 80% of the D15 protein remained soluble. This soluble rD15 antigen was used for the immunogenicity studies described below. From shake-flask experiments, it was estimated that about 10 mg of soluble rD15 protein was obtained from 1 L of E. coli bacterial culture. It is clear that growing the recombinant E. coli strains under optimised fermentation conditions significantly increase the level of rD15 production.

5
10 (vi) Immunogenicity of the full-length recombinant D15 protien (rD15).

The immunogenicity of the full-length rD15 protein was studied in guinea pigs and mice. Using the immunization protocols described in Figure 7, a 15 µg dose of rD15 induced high IgG titers in guinea pigs when administered in the presence of either Freund's adjuvant or AlPO₄. In the mouse dose-response study, the protein appeared to be immunogenic at a dose as low as 5 µg in either Freund's adjuvant (Figure 8A) or AlPO₄ (Figure 8B).

20 The protective ability of rD15 against H. influenzae type b infection was examined in the infant rat model of bacteremia essentially as described by Loeb (1987). Thus, infant rats passively immunized with guinea pig anti-rD15 antisera were significantly less bacteremic than controls injected with pre-bleed sera, which is consistent with the previous report by Thomas et al. (1990).

25 (vii) Purification and characterization of the N-terminal rD15 fragment.

30 The truncated rD15 fragment corresponding to the N-terminus of the D15 protein (residues 22 to 223) as described in Example 6, was expressed in E. coli as a soluble protein fused to GST. The fusion protein (46 kDa) was readily extracted using phosphate buffered saline (PBS). Purification of the GST-D15 fragment fusion

the immunization protocols described in Figure 10, a 10 μ g dose of N-terminal rD15 fragment induced a good booster response in guinea pigs with almost all the adjuvants tested. The highest anti-D15 IgG titer was observed in the group of guinea pigs immunized with N-terminal rD15 fragment in Freund's adjuvant. The second best adjuvant was Titermax (CytRx Inc.). The other two adjuvants, TPAD4 (tripalmityl-Cys-Ser-Glu₄) and AlPO₄ were equally potent.

5 (ix) **Protective ability of the N-terminal rD15 fragment against H. influenzae type b challenge.**

An in vivo challenge model for assessing the protective abilities of antigen against diseases caused by Haemophilus is the infant rat model of bacteremia as described by Loeb 1987. The protective ability of the N-terminal rD15 fragment against H. influenzae type b challenge was examined in this rat model. As illustrated in Table 1, infant rats passively immunized with rabbit anti-N-terminal rD15 fragment antisera showed significantly lower bacteremia compared to those injected with pre-bleed sera.

Since passively transferred antisera against the N-terminal rD15 fragment were found to be protective in the infant rat model of bacteremia, it was of interest to identify the protective epitope(s) of this N-terminal rD15 fragment. The first nine overlapping peptides of the D15 protein as listed in Table 2 were chemically synthesized based upon the amino acid sequence derived from the sequence of the D15 gene from H. influenzae type b Ca (Figure 1). These synthetic peptides were assessed for their reactivities with either rabbit or guinea pig antisera raised against purified N-terminal rD15 fragment by ELISAs. As shown in Table 3, both guinea pig and rabbit antisera reacted with a cluster of D15 peptides, including peptides D15-P4 to D15-P8 encompassing residues 93 to 209 of the D15 primary sequence.

was completely blocked by the addition of this mixture of five peptides (Table 5, group #2, 106%, $p = 0.53 \times 10^{-8}$). These results strongly indicate that a cocktail of D15 synthetic peptides may be used as immunogens to induce protective antibodies against H. influenzae.

(x) Epitope prediction and peptide synthesis.

To map the immunodominant T-cell or B-cell epitopes of D15, overlapping synthetic peptides covering the entire D15 protein sequence (Table 2 - SEQ ID NO: 14 to 49) were synthesized using the t-Boc solid-phase peptide synthesis as described in Example 15. The peptides were chosen based on their high index of hydrophilic β -turns estimated by secondary structure prediction analysis (Figure 11). Such peptides are likely to be surface-exposed and antigenic. Peptides more than 25 residues in length were selected to better mimic native epitopes.

(xi) Identification and characterization of immunodominant epitopes of D15 using synthetic peptides.

To map the linear B-cell epitopes of D15, overlapping synthetic peptides representing the entire sequence of D15 were individually coated onto ELISA plates and probed with several anti-rD15 antisera as described in Example 19. The results are summarized in Table 6. Mouse antisera raised against rD15 reacted with all D15 peptides, but the major epitopes were located within peptides D15-P8 (residues 180-209 - SEQ ID NO: 21), D15-P10 (residues 219-249 - SEQ ID NO: 23), D15-P11 (residues 241-270 - SEQ ID NO: 24), and D15-P26 (residues 554-582 - SEQ ID NO: 39), respectively. Rabbit anti-D15 antisera recognized only peptides D15-P4 (residues 93-122 - SEQ ID NO: 17), D15-P14 (residues 304-333 - SEQ ID NO: 27) and D15-P36 (residues 769-798 - SEQ ID NO: 49). Guinea pig antisera raised against rD15 reacted with peptides D15-P2 (residues 45-72 - SEQ ID NO: 15), D15-P4 (residues 93-122 - SEQ ID NO: 17), D15-P6 (residues 135-164 - SEQ ID NO: 19), D15-P8 (residues 180-209 - SEQ ID

cells in the immune system is to provide helper activity for eliciting high levels of antigen-specific antibodies following immunization. Antigens containing Th1 epitope(s) stimulate antigen-specific T-cells to produce
5 high levels of IL-2 and IFN- γ , whereas Th2 epitope(s) induce high levels of IL-4 expression. Th0 epitope(s) stimulate the synthesis of IFN- γ and IL-4.

Little is known about the cellular immune response to outer membrane proteins of H. influenzae and its role
10 in the protection against H. influenzae infection and diseases. To this end, the inventors performed studies of the cellular response elicited in mice following rD15 immunization. D15-specific T-cell epitopes were determined using D15 peptides and T-cell lines obtained
15 from five BALB/c mice immunized with rD15 (see Example 23). The lymphocyte proliferative responses of the D15-specific T-cell lines to overlapping D15 peptides were determined in conventional cytokine assays as described in Example 24. The results summarized in Table 7,
20 revealed that stimulation only with certain synthetic peptides elicited proliferative responses and the release of specific cytokines. Synthetic peptides corresponding to residues 114-143 (D15-P5 - SEQ ID NO: 18), 282-312 (D15-P13 - SEQ ID NO: 26) and 577-602 (D15-P27 - SEQ ID
25 NO: 40), and 219-249 (D15-P10 - SEQ ID NO: 23), 262-291 (D15-P12 - SEQ ID NO: 25), 390-416 (D15-P18 - SEQ ID NO: 31), 410-435 (D15-P19 - SEQ ID NO: 32) 554-582 (D15-P26 - SEQ ID NO: 39), 596-625 (D15-P28 - SEQ ID NO: 41), 725-750 (D15-P34 - SEQ ID NO: 47) and 745-771 (D15-P35 - SEQ
30 ID NO: 48) were shown to be highly stimulatory for rD15-specific BALB/c Th0 cells and Th1 cells, respectively. Therefore, these immunodominant T-cell epitopes can be used as autologous carriers for PRP, and/or OMP B-cell epitopes to enhance their immunogenicity. The Th1 cell
35 epitopes identified above may be useful in the H.

linked to polysaccharides including PRP as synthetic glycopeptide or lipoglycopeptide conjugates to produce alternate vaccines. These vaccines can be used to immunize against diseases caused by H. influenzae when administered to mammals, for example, by the intramuscular or parenteral route, or when delivered using microparticles, capsules, liposomes and targeting molecules, such as toxins or fragments thereof, and antibodies, to cells of the immune system or mucosal surfaces.

(xiv) Utility of D15 as carrier protein for the production of glycoconjugates.

To determine whether D15 may serve both as a protective antigen and a carrier, D15-PRP conjugation experiments were performed as described in Example 14. The D15-PRP conjugates were found to be highly immunogenic in rabbits and able to elicit both anti-D15 and anti-PRP IgG antibody responses as judged by D15-specific ELISA and PRP-BSA immunoassay (Table 9). These results clearly demonstrate the practical utility of D15 as a carrier protein for glycoconjugation technology.

In preferred embodiments of the present invention, the carrier function of D15 can be generally utilized to prepare chimeric molecules and conjugate vaccines against pathogenic bacteria, including encapsulated bacteria. Thus, the glycoconjugates of the present inventions may be applied to vaccinations to confer protection against infection with any bacteria having polysaccharide antigens, including, for example, Haemophilus influenzae, Streptococcus pneumoniae, Escherichia coli, Neisseria meningitidis, Salmonella typhi, Streptococcus mutans, Cryptococcus neoformans, Klebsiella, Staphylococcus aureus and Pseudomonas aeruginosa.

In another embodiment, the carrier function of D15 may be used, for example, to induce immunity toward abnormal polysaccharides of tumor cells, or to produce

rD15 protein and its fragments were found to cross-react immunologically with the native D15 antigen isolated from both typeable and non-typeable H. influenzae isolates and thus represent cross-reactive immunogens for inclusion in a vaccine against diseases caused by H. influenzae.
5 Furthermore, Haemophilus convalescent serum recognized D15 purified from H. influenzae as described herein, rD15 and N-terminal rD15 fragment.

In another embodiment, the present invention provides a gene coding for the outer membrane protein D15 from H. influenzae having the specific nucleotide sequences described herein or ones substantially homologous thereto (i.e. those which hybridize under stringent conditions to such sequences), for genetically engineering hybrids or chimeric proteins containing a D15
15 fragment fused to another polypeptide or protein or a polysaccharide, such as H. influenzae outer membrane proteins, for example, P1, P2, or P6 or PRP. As a result, the hybrids, chimeric proteins or glycoconjugates may have higher protectivity against H. influenzae than
20 D15, or P1, or P2, or P6, or PRP alone.

Thus, D15 outer membrane protein can function both as a protective antigen and as a carrier in a conjugate vaccine to provide autologous T-cell priming, wherein the
25 hapten part of the conjugate is the capsular polysaccharide moiety (PRP) of H. influenzae. This D15-carbohydrate conjugate can elicit antibodies against both PRP and D15, and thus should enhance the level of protection against H. influenzae-related diseases,
30 especially in infants.

In another embodiment, the present invention comprises an essentially pure form of at least one protein or peptide containing an amino acid sequence corresponding to at least one antigenic determinant of
35 D15, which peptide is capable of eliciting polyclonal antibodies against H. influenzae in mammals. These D15-

catarrhalis, Staphylococcus aureus, or respiratory syncytial virus, in the presence or absence of adjuvant.

The D15 peptides (Table 2) or any portion, variant or mutant thereof, can easily be synthesized either
5 manually or with a commercially available peptide synthesizer, such as the Applied Biosystems Model 430A synthesizer.

It is clearly apparent to one skilled in the art, that the various embodiments of the present invention
10 have many applications in the fields of vaccination, diagnosis, and treatment of diseases caused by Haemophilus infections, and the generation of immunological reagents. A further non-limiting discussion of such uses is further presented below.

15 1. Vaccine preparation and use

Immunogenic compositions, suitable for use as vaccines, may be prepared from immunogenic D15 outer membrane protein, fragments or analogs thereof and/or peptides corresponding to portions of D15 as disclosed
20 herein. The vaccine elicits an immune response which produces antibodies, including anti-D15 outer membrane protein antibodies and antibodies against D15 that are opsonizing or bactericidal. Should the vaccinated subject be challenged by Haemophilus, the antibodies bind
25 to the D15 outer membrane protein and thereby inactivate the bacterium. Opsonizing and bactericidal antibodies represent examples of antibodies useful in protection against disease.

Vaccines containing peptides are generally well
30 known in the art, as exemplified by U.S. Patents 4,601,903; 4,599,231; 4,599,230; and 4,596,792; all of which references are incorporated herein by reference. As to any further reference to patents and references in this description, they are as well hereby incorporated by
35 reference without any further notice to that effect. Vaccines may be prepared as injectables, as liquid

However, suitable dosage ranges are readily determinable by one skilled in the art and may be of the order of micrograms of the D15 outer membrane protein, analog, fragment and/or peptides. Suitable regimes for initial
5 administration and booster doses are also variable, but may include an initial administration followed by subsequent administrations. The dosage of the vaccine may also depend on the route of administration and varies according to the size of the host.

10 The nucleic acid molecules encoding the D15 outer membrane protein of the present invention may also be used directly for immunization by administration of the DNA directly, for example, by injection for genetic immunization or by constructing a live vector, such as
15 Salmonella, BCG, adenovirus, poxvirus or vaccinia. A discussion of some live vectors that have been used to carry heterologous antigens to the immune system are discussed in, for example, O'Hagan (1992). Processes for the direct injection of DNA into test subjects for
20 genetic immunization are described in, for example, Ulman et al. (1993).

The use of peptides in vivo may first require their chemical modification since the peptides themselves may not have a sufficiently long serum and/or tissue half-
25 life. Such chemically modified peptides are referred to herein as peptide analogs. The term peptide analog extends to any functional chemical equivalent of a peptide characterized by its increased stability and/or efficacy in vivo or in vitro in respect of the practice
30 of the invention. The term peptide analog is also used herein to extend to any amino acid derivative of the peptides as described herein. Peptide analogs contemplated herein are produced by procedures that include, but are not limited to, modifications to side
35 chains, incorporation of unnatural amino acids and/or their derivatives during peptide synthesis and the use of

nitration with tetranitromethane to form a 3-nitrotyrosine derivative.

Modification of the imidazole ring of a histidine residue may be accomplished by alkylation with iodoacetic acid derivatives or N-carbethoxylation with diethylpyrocarbonate.

Examples of incorporating unnatural amino acids and derivatives during peptide synthesis include, but are not limited to, use of norleucine, 4-amino butyric acid, 4-amino-3-hydroxy-5-phenylpentanoic acid, 6-aminohexanoic acid, t-butylglycine, norvaline, phenylglycine, ornithine, sarcosine, 4-amino-3-hydroxy-6-methylheptanoic acid, 2-thienyl alanine and/or D-isomers of amino acids.

2. Immunoassays

The D15 outer membrane protein, analog, fragment and/or peptides of the present invention are useful as antigens in immunoassays, including enzyme-linked immunosorbent assays (ELISA), RIAs and other non-enzyme linked antibody binding assays or procedures known to the art for the detection of anti-bacterial, Haemophilus, D15 and/or peptide antibodies. In ELISA assays, the D15 outer membrane protein, fragment or analogs thereof and/or peptides corresponding to portions of D15 outer membrane protein are immobilized onto a selected surface, for example, a surface exhibiting a protein affinity, such as the wells of a polystyrene microtiter plate. After washing to remove incompletely adsorbed D15 outer membrane protein, analog, fragment and/or peptides, a nonspecific protein, such as bovine serum albumin (BSA) or casein, that is known to be antigenically neutral with regard to the test sample may be bound to the selected surface. This allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus decreases the background caused by nonspecific bindings of antisera onto the surface. Normally, the peptides

Haemophilus and other bacteria that have genes encoding D15 outer membrane proteins.

5 The nucleotide sequences comprising the sequence encoding the D15 outer membrane protein of the present invention are useful for their ability to selectively form duplex molecules with complementary stretches of other D15 genes. Depending on the application, a variety of hybridization conditions may be employed to achieve varying degrees of selectivity of the probe toward the
10 other D15 genes. For a high degree of selectivity, stringent conditions are used to form the duplexes, such as low salt and/or high temperature conditions, such as provided by 0.02 M to 0.15 M NaCl at temperatures of between about 50°C to 70°C. For some applications, less
15 stringent hybridization conditions are required such as 0.15 M to 0.9 M salt, at temperatures ranging from between about 20°C to 55°C. Hybridization conditions can also be rendered more stringent by the addition of increasing amounts of formamide, to destabilize the
20 hybrid duplex. Thus, particular hybridization conditions can be readily manipulated, and will generally be a method of choice depending on the desired results.

In a clinical diagnostic embodiment, the nucleic acid sequences of the D15 outer membrane protein genes of
25 the present invention may be used in combination with an appropriate means, such as a label, for determining hybridization. A wide variety of appropriate indicator means are known in the art, including radioactive, enzymatic or other ligands, such as avidin/biotin, which
30 are capable of providing a detectable signal. In some diagnostic embodiments, an enzyme tag, such as urease, alkaline phosphatase or peroxidase, instead of a radioactive tag may be used. In the case of enzyme tags, colorimetric indicator substrates are known which can be
35 employed to provide means visible to the human eye or

The pBR322 plasmid, or other microbial plasmid or phage must also contain, or be modified to contain, promoters which can be used by the microbial organism for expression of its own proteins.

5 In addition, phage vectors containing replicon and control sequences that are compatible with the host microorganism can be used as a transforming vector in connection with these hosts. For example, the phage in
10 lambda GEM™-11 may be utilized in making recombinant phage vectors which can be used to transform host cells, such as E. coli LE392.

 Promoters commonly used in recombinant DNA construction include the β -lactamase (penicillinase) and lactose promoter systems and other microbial promoters,
15 such as the T7 promoter system. Details concerning the nucleotide sequences of promoters are known, enabling a skilled worker to ligate them functionally with plasmid vectors. The particular promoter used generally is a matter of choice depending upon the desired results.
20 Hosts that are appropriate for expression of the transferrin receptor genes, fragment analogs or variants thereof include E. coli, Bacillus, Haemophilus, Bordetella, fungi, yeast, or the baculovirus and poxvirus expression systems may be used.

25 In accordance with an aspect of this invention, it is preferred to make the D15 outer membrane protein, fragment or analog thereof by recombinant methods, particularly since the naturally occurring D15 protein as purified from culture of a species of Haemophilus may
30 include undesired contaminants, including trace amounts of toxic materials. This problem can be avoided by using recombinantly produced D15 outer membrane protein in heterologous systems which can be isolated from the host in a manner to minimize toxins in the purified material.
35 Particularly desirable hosts for expression in this regard include Gram positive bacteria which do not have

the invention. Although specific terms have been employed herein, such terms are intended in a descriptive sense and not for purposes of limitations. Immunological and recombinant DNA methods may not be explicitly described in this disclosure but are well within the scope of those skilled in the art.

EXAMPLES

Methods of molecular genetics, protein biochemistry, and immunology used but not explicitly described in this disclosure and these EXAMPLES are amply reported in the scientific literature and are well within the ability of those skilled in the art.

Example 1

This Example illustrates the cloning and sequencing of the D15 genes.

Genomic DNA was purified from the Haemophilus influenzae type b strain Ca by lysis of the bacteria with pronase and sodium dodecylsulphate followed by phenol extraction and isopropanol precipitation, according to Berns and Thomas, 1965. The DNA was then partially digested with EcoRI and the DNA fraction containing 6-10 kb fragments was isolated following electrophoresis in low-melting point agarose. These fragments were ligated into a lambda gt11 Amp^r vector (Thomas and Rossi, 1986) and cloned as a lysogen into E. coli strain BTA282. Recombinant clones were selected for their ampicillin resistance conferred by the vector. To identify clones producing H. influenzae type b antigen, the clones were replica-plated on nitrocellulose filters and duplicate colonies induced for expression by temperature switch to 42°C for 2 hours. Colonies were lysed by wetting the filters with 1% sodium dodecylsulphate (SDS). The filters were then placed into a chloroform-saturated atmosphere for 15 min. The filters were then assayed by colony radioimmuno-assay using a hyperimmune rabbit anti-H. influenzae type b antiserum absorbed with E. coli lysate

plasmid transformed into E. coli HB101. Recombinant bacteria were found to produce the expected M_r 80 kDa H. influenzae type b antigen when examined by Western blotting. The insert DNA was then characterised by restriction endonuclease mapping. A 2.8 kb HindIII-EcoRI fragment was subcloned into pUC19 to generate plasmid pUC19/D15, which was transformed into E. coli HB101. The recombinant bacteria expressed a M_r 80 kD protein recognized by D15-specific antibodies on Western blot analysis of E. coli lysates.

Plasmid DNA was prepared from two individual colonies of recombinant E. coli HB101 containing the pUC19/D15 plasmid using standard techniques. Oligonucleotide sequencing primers of 17-25 bases in length were synthesized on the ABI model 380B DNA Synthesizer and purified by chromatography using OPC cartridges obtained from Applied Biosystems Inc., and used in accordance with the manufactures recommendations. Samples were sequenced using the ABI model 370A DNA Sequencer and dye terminator chemistry according to manufacturers' protocols. This sequence analysis indicated that the D15 gene contains an open reading frame encoding for 789 amino acids, including a putative signal sequence (Figure 1). The derived amino acid sequence was found to contain the sequence of an internal peptide obtained by thrombin digestion of native D15 that had been chemically determined. The amino acid composition of D15 derived from the D15 gene sequence was comparable (within experimental error) to that of the native protein as determined by amino acid analysis.

Example 2

This Example illustrates the preparation of chromosomal DNA from Haemophilus influenzae strains Eagan, MinnA, SB33, and PAK 12085.

H. influenzae Eagan and PAK 12085 chromosomal DNAs were digested with Sau3A I (0.5 unit/10 µg DNA) at 37°C for 15 minutes and size-fractionated by agarose gel electrophoresis. Gel slices corresponding to DNA fragments of 15-23 kb were excised and DNA was electroeluted overnight in dialysis tubing containing 3 mL of TAE (40mM Tris-acetate, 1mM EDTA, pH 8.0) at 14V. The DNA was precipitated twice and resuspended in water before overnight ligation with EMBL3 BamH I arms (Promega). The ligation mixture was packaged using the Lambda in vitro packaging kit (Amersham) according to the manufacturer's instructions and plated onto E. coli NM539 cells. The library was titrated, then amplified and stored at 4°C under 0.3% chloroform.

Minna chromosomal DNA (10 µg) was digested with Sau3A I (40 units) for 2, 4, and 6 minutes then size-fractionated on a 10-30% sucrose gradient in TNE (20mM Tris-HCl, 5mM NaCl, 1mM EDTA, pH 8.0). Fractions containing DNA fragments >5 kb were pooled and precipitated. In a second experiment, chromosomal DNA (2.6 µg) was digested with Sau3A I (4 units) for 1, 2, and 3 minutes and size-fractionated by preparative agarose gel electrophoresis. Gel slices containing DNA fragments of 10-20 kb were excised and DNA extracted by a standard freeze/thaw technique. The size-fractionated DNA from the two experiments was pooled for ligation with BamH I arms of EMBL3 (Promega). The ligation mix was packaged using the Gigapack II packaging kit (Amersham) and plated on E. coli LE392 cells. The library was titrated, then amplified and stored at 4°C under 0.3% chloroform.

SB33 chromosomal DNA (20 µg) was digested with Sau3A I (40 units) for 2, 4, or 6 minutes and size-fractionated on a 10-30% sucrose gradient in TNE (20mM Tris-HCl, 5mM NaCl, 1mM EDTA, pH 8.0). Fractions containing fragments >5 kb were pooled. In a second experiment, SB33

compared with the amino acid sequence of the D15 protein of H. influenzae type b Ca (Figure 3).

Example 5

5 This Example illustrates the expression of rD15 protein in E. coli.

A 2.8 kb fragment HindIII-EcoRI was subcloned into pUC19 and this pUC19/D15 plasmid was transformed into E. coli HB101. Upon induction, the positive clones expressed an 80 kDa protein which was recognized by D15-specific
10 antisera on Western blot analysis. A HindIII-Pst I fragment was also subcloned into pUC19 and shown to express a 67 kDa protein. According to the restriction map, this 67 kDa protein corresponded to a C-terminal truncated D15 protein. On Western blot analysis, this
15 truncated D15 was still recognized by the D15-specific antisera.

Plasmids to express the D15 gene of the non-typeable strain SB33 in E. coli were constructed. Plasmid JB-1042-5-1 containing the SB33 D15 gene and its flanking
20 regions, was digested with EcoR I and Hind III and the 3kb D15 insert subcloned into pUC to give plasmid pRY-60-1 (Figure 4). Appropriate oligonucleotides were synthesized to restore the native D15 sequence between the ATG codon of the expression plasmid pT7-7 and the
25 BsrF I site within the D15 gene. These oligonucleotides had the following sequence:

Nde

30 5' - TATGGCACCTTTTGTGGCAAAGATATTCGTGTGGATGGTGTTC AAGGTG
ACCGTGGAAAACACCGTTTTCTATAAGCACACCTACCACAAGTTCCACTGAATCT
ACTTAGAATCAACAAACCGAGCAAGTTTACCTGTTTCGTG - SEQ ID NO: 50
35 TGGTTGTTTAGGCTCGTTCAAATGGACAAGCACGGCC-5' - SEQ ID NO: 51
BsrF I

Plasmid pRY-60-1 was digested with EcoR I and BsrF I and the DNA fragment containing most of the D15 gene was
40 purified. pUC was digested with EcoR I and Nde I and the

produced by transformed E. coli was isolated by affinity purification on glutathione agarose.

Example 7

5 This Example describes alternative expression systems for rD15.

The D15 gene or fragments thereof are also expressed in E. coli under the control of other regulated promoters. The D15 gene or fragments thereof are expressed in the absence of the leader peptide, or in
10 other cloning systems where toxicity of D15 expression to the host is not problematic. The gene or fragments thereof are synthesized de novo or by employing the polymerase chain reaction using suitable primers. These genes are cloned into suitable cloning vectors or
15 bacteriophage vectors in E. coli or other suitable hosts directly when toxicity can be avoided. Expression systems are Gram-positive bacteria (such as Bacillus species), pox virus, adenovirus, baculovirus, yeast, fungi, BCG or mammalian expression systems.

20 Example 8

This Example illustrates the protocol for extraction and purification of rD15 from E. coli expression system.

The cell pellet from a 250 mL culture, prepared as described in Example 5, was resuspended in 40 mL of 50 mM
25 Tris, pH 8.0, and disrupted by sonication (3 x 10 min, 70% duty circle). The extract was centrifuged at 20,000 x g and the resulting pellet saved. The initial pellet was re-extracted with 40 mL of 50 mM Tris, 0.5% Triton X-100, 10 mM EDTA, pH 8.0. The suspension was then
30 sonicated for 10 minutes at 70% duty circle. The extract was centrifuged at 300 x g for 5 minutes. The resulting supernatant was centrifuged again at 20,000 x g for 30 min and the resulting pellet was saved. The pellet was resuspended in 50 mM Tris, 0.5% Triton X-100, 10 mM EDTA,
35 pH 8.0. The suspension was then mixed with PBS/ 8 M urea to a final urea concentration of 6 M. The solution was

This Example illustrates the procedure used for N-terminal rD15 fragment purification from GST using Glutathione-Sepharose 4B affinity chromatography.

5 A thrombin-digested GST-(D15 fragment) sample, prepared as described in Example 10, was loaded onto a Glutathione-Sepharose 4B column (2 mL) equilibrated with PBS containing 1% Triton X-100. The run-through of the column containing the N-terminal rD15 fragment was saved. After washing the column with 20 mL of PBS, the affinity
10 column was regenerated by removing GST using 50 mM Tris-HCl buffer, pH 8.0, containing 5 mM glutathione. The purity of rD15 fragment was analysed by SDS-PAGE (Figure 9, lane 5). This N-terminal rD15 fragment contains amino acids 63-223 of the D15 protein as a result of cleavage
15 at the spacious thrombin site shown in Figure 1A.

Example 12

This Example illustrates the protocol used for the purification of D15-specific polyclonal antibodies by affinity chromatography using GST-(D15 fragment) fusion
20 protein.

The recombinant GST-(D15 fragment) fusion protein, prepared as described in Example 9, was conjugated to cyanogen bromide-activated Sepharose. The affinity column was then used to purify antibodies from a rabbit
25 hyperimmune anti-H. influenzae type b antiserum. The affinity purified-antibodies were shown by immunoblotting to react with a 80 kDa component present in the lysates of E. coli transformed with pUC9/D15 and in the lysates of several typeable and nontypeable H. influenzae
30 isolates. These results confirmed that the DNA segment encoding the D15 fragment of the fusion protein was part of the open reading frame of the D15 gene.

Similarly, antisera raised against the recombinant fusion protein (Example 9) or the purified N-terminal
35 rD15 fragment (Example 11) reacted with the D15 protein produced by H. influenzae strains (Example 13).

in Example 17. The mean molecular size of the PRP molecules used for conjugation was determined as being approximately 20,000 Daltons. The conjugation was carried out without a linker molecule but may also be carried out with a linker molecule. A PRP/D15 molar ratio of approximately 7 was used to provide an excess of PRP hapten.

The PRP/rD15 conjugate was tested according to the protocol of Example 18 for immunogenicity in rabbits and elicited both primary and secondary anti-PRP IgG and anti-D15 antibody responses (Table 9). Rabbit anti-rD15-PRP antisera also strongly reacted with both native D15 and rD15 as judged by immunoblot analysis. These data indicate that rD15 can be used as a carrier protein in a conjugate vaccine. In addition, a rD15-PRP conjugate vaccine should ensure a more consistent protection against H. influenzae type b disease, particularly in infants, as a result of the additional homotypic protection provided by antibodies directed against the D15 protein.

Example 15

This Example describes the preparation of D15 peptides.

D15 peptides (Table 2) were synthesized using an ABI 430A peptide synthesizer and optimized t-Boc chemistry as described by the manufacturer, then cleaved from the resin by hydrofluoric acid (HF). The peptides were purified by reversed-phase high performance liquid chromatography (RP-HPLC) on a Vydac C4 semi-preparative column (1 x 30 cm) using a 15 to 55% acetonitrile gradient in 0.1% trifluoryl acetic acid (TFA) developed over 40 minutes at a flow rate of 2 mL/min. All synthetic peptides (Table 2) used in biochemical and immunological studies were >95% pure as judged by analytical HPLC. Amino acid composition analyses of

equilibrated with 0.2 M sodium phosphate buffer, pH 7.2, and eluted with the same buffer. Fractions were monitored for absorbance at 230 nm. The first major protein peak was pooled and concentrated in a Centriprep 30 to 2.2 mL.

- 5 The amount of protein was determined using the Bio Rad protein assay, and was found to be 300 µg/mL. The presence of PRP in the protein conjugate fraction was confirmed by the Orcinol test.

Example 18

- 10 This Example describes the protocol used for the production of anti-PRP antisera in animals using rD15-PRP conjugates.

Rabbits were immunized intramuscularly with rD15-PRP conjugates (Example 14) (5 to 50 µg PRP equivalent) mixed
15 with 3 mg AlPO₄ per mL, followed by two booster doses (half amount of the same immunogen) at 2 week intervals. Antisera were collected every 2 weeks after the first injection, heat-inactivated at 56°C for 30 minutes and stored at -20°C.

20 Example 19

This Example illustrates the reactivity between D15 peptides and anti-peptide and D15-specific antisera using D15-specific and peptide-specific ELISAs.

- Microtiter wells (Nunc-Immunoplate, Nunc, Denmark)
25 were coated with 200 ng of purified rD15 or 500 ng of individual peptides in 50 µL of coating buffer (15 mM Na₂CO₃, 35 mM NaHCO₃, pH 9.6) for 16 hours at room temperature. The plates were then blocked with 0.1% (w/v) BSA in phosphate buffer saline (PBS) for 30 minutes at
30 room temperature. Serially diluted antisera were added to the wells and incubated for 1 hour at room temperature. After removal of the antisera, the plates were washed five times with PBS containing 0.1% (w/v) Tween-20 and 0.1% (w/v) BSA. F(ab')₂ fragments from goat anti-rabbit,
35 guinea pig, mouse, or human IgG antibodies conjugated to horseradish peroxidase (Jackson ImmunoResearch Labs Inc.,

the substrate tetramethylbenzidine (TMB) in H₂O₂ (ADI, Toronto). The reaction was stopped with 1N H₂SO₄ and the optical density measured at 450 nm using a Titretrek Multiskan II (Flow Labs., Virginia). A standard anti-PRP antiserum of known titer was included as positive control. Assays were performed in triplicate, and the reactive titer of each antiserum was defined as the reciprocal of the dilution consistently showing a 2-fold increase in O.D. value over that obtained with the pre-immune serum (Table 9).

Example 21

This Example describes the protocol used for the production of D15-specific antisera using purified D15, rD15 or N-terminal rD15 fragment.

New Zealand White rabbits (Maple Lane) and guinea pigs (Charles River) were immunized intramuscularly (IM) with a 10 µg dose of either affinity-purified native D15 (Example 13), recombinant D15 (Example 8) or N-terminal rD15 fragment (Example 11) emulsified in Freund's complete adjuvant (Difco). Animals were boosted on day 28 with another 10 µg dose of affinity-purified D15 or rD15 or rD15 fragment emulsified in Freund's incomplete adjuvant and bled on day 42 via the marginal ear vein. D15-specific polyclonal antibodies were purified from this material as described in Example 12.

Example 22

This Example illustrates the protective activity of D15-specific antisera against H. influenzae type b challenge using the infant rat model of bacteremia.

Five-day old infant rats were inoculated subcutaneously (SC) on the dorsum with 0.15 mL of two different rabbit anti-N-terminal rD15 fragments. Pre-immune sera were used as negative controls. One day after immunization, the infant rats were injected intraperitoneally (IP) with 200 colony-forming units (cfu) of Haemophilus influenzae type b Minn A strain (0.1

supernatant added to further expand and maintain the viability of the peptide-specific T-cells. After a further 6 day-incubation, the cells were washed three times, each time with 200 μ L of culture medium.

5 Each set of cultures was then stimulated with the corresponding concentrations (1, 10 and 100 μ g per mL) of the peptide in the presence of 2×10^5 irradiated (1,500 rad) BALB/c spleen cells in a final volume of 200 μ L of culture medium. Sixty μ L of supernatant were then removed
10 from each microculture. The supernatants from each triplicate cultures set were pooled. All supernatants were assayed for IL-2, Interleukin-4 and Interferon-gamma (IFN- γ). Detections of IL-2 and IL-4 were performed using murine IL-2 and IL-4 ELISA kits purchased from
15 Endogen Inc. (MA, USA) respectively. Assay of IFN- γ was performed using a mouse IFN- γ ELISA kit supplied by Genzyme Corporation (MA, USA). Test culture supernatants were assayed at 1 in 5 dilution according to the manufacturers' instructions. The results obtained are
20 set forth in Table 7.

Example 25

This Example describes the general procedure used for the production of murine D15-specific monoclonal antibodies.

25 BALB/c mice were immunized intraperitoneally with 20 to 50 μ g of the N-terminal rD15 fragment (Example 11) emulsified in Freund's complete adjuvant. Two weeks later, the mice were given another injection of the same amount of immunogen in incomplete Freund's adjuvant
30 (IFA). Three days before the fusion, the mice were boosted again with the same amount of immunogen in IFA. Hybridomas were produced by fusion of splenic lymphocytes from immunized mice with non-secreting Sp2/0 myeloma cells as previously described by Hamel et al. (1987).
35 D15-specific hybridomas were cloned by sequential limiting dilutions and screened for anti-D15 monoclonal

TABLE 1

PROTECTIVE EFFECT OF PASSIVELY TRANSFERRED ANTI-N-TERMINAL RD15
FRAGMENT ANTIBODIES IN THE INFANT RAT MODEL OF BACTEREMIA¹

Rabbit antisera	cfu/0.1 mL blood		p value
	Pre-immune	Post-immunization	
Rb#434	510 (6/6) ²	6 (1/6)	<0.001
Rb#435	910 (4/4)	6 (1/4)	<0.001

¹ Five-day old infant rats were passively immunized with 0.15 mL of rabbit anti-N-terminal rD15 fragment s.c. One day later, the infant rats were challenged with 200 cfu of H. influenzae type b strain Minna (0.1 mL, IP). The blood samples were taken from each rat 24 hours after the challenge and analysed for bacteria counts.

² The parentheses indicate the number of rats found to be bacteremic out of the total number of rats challenged.

D15-P29	619-646	LWVVSASAGYANGFGNKRLPFYQTYT	42
D15-P30	641-666	FYQTYTAGGIGSLRGFAYGSIGPNAI	43
D15-P31	662-688	GPNAIYAEYGNNGSGTGTFKKISSDVIG	44
D15-P32	681-709	KISSDVIGGNALATASAELIVPTPFVSDK	45
D15-P33	705-731	FVSDKSQNTVRTSLFVDAASVWNTKWK	46
D15-P34	725-750	VWNTKWKSDKNGLSDVLKRLPDYGK	47
D15-P35	745-771	LPDYGKSSRIRASTGVGFQWQSPIGPL	48
D15-P36	769-798	GPLVFSYAKPIKKYENDDVEQFQFSIGGSF	49

TABLE 4

INHIBITION OF ANTI-N-TERMINAL rD15 FRAGMENT ANTIBODY-INDUCED
PROTECTION BY D15 PEPTIDES IN THE INFANT RAT MODEL OF BACTEREMIA

Group #	Antibody	cfu / 10 μ L blood	cfu in each group/ cfu in group #4 (control) (%)
1	Anti-D15 Ab + PBS	60 \pm 120 (3/7)	3
2	Anti-D15 Ab + peptides	300 \pm 240 (6/7)	13
3	Anti-D15 Ab + rD15	1,520 \pm 1,280 (7/7)	64
4	PBS + peptides	2,360 \pm 1,200 (6/7)	100

One half mL of rabbit anti-N-terminal rD15 fragment antiserum (Anti-rD15 fragment Ab) was mixed with either nine D15 peptides (100 μ g of peptides D15-P2 to D15-P10, See TABLE 2) or with 600 μ g of N-terminal rD15 fragment at room temperature for 1 hr. Antiserum and peptides mixed with PBS were used as controls. Seven-day old infant rats were injected s.c. with 0.2 mL of the various preparations. After 24 h, the infant rats were challenged I.P. with 200 cfu of *H. influenzae* type b strain MinnA. The blood samples were taken at 24 h after the challenge. The numbers in parentheses indicate the number of animals that were bacteremic out of the total number of animals challenged. The level of bacteremia is expressed as the mean of values obtained from seven infant rats tested individually \pm one standard deviation (SD).

TABLE 6

REACTIVITY OF RABBIT, GUINEA PIG AND MOUSE ANTI-rD15 ANTISERA
WITH D15 PEPTIDES

Peptide	Rabbit ²	Reactive Titer ¹	
		Guinea Pig ³	Mouse ⁴
D15-P1	-	-	+
D15-P2	-	+++	+
D15-P3	-	-	+
D15-P4	+	+	+
D15-P5	-	-	+
D15-P6	-	+	+
D15-P7	-	-	+
D15-P8	-	++++	++++
D15-P9	-	-	+
D15-P10	-	-	+++
D15-P11	-	-	+++
D15-P12	-	-	+
D15-P13	-	-	+
D15-P14	+++	+	+
D15-P15	-	-	+
D15-P16	-	-	+
D15-P17	-	-	+
D15-P18	-	-	+
D15-P19	-	-	+
D15-P20	-	-	+
D15-P21	-	-	+
D15-P22	-	-	+
D15-P23	-	-	+
D15-P24	-	-	+
D15-P25	-	-	+
D15-P26	-	-	+++
D15-P27	-	+	+

TABLE 7

T-CELL STIMULATORY ACTIVITY OF D15 PEPTIDES

Peptide	IL-2 ²	CYTOKINE RELEASE (pg/mL) ¹	
		γ -IFN ³	IL-4 ⁴
D15-P1	-	-	-
D15-P2	122	-	-
D15-P3	25	-	-
D15-P4	-	-	-
D15-P5	742	38,000	13
D15-P6	-	-	-
D15-P7	-	-	-
D15-P8	-	-	-
D15-P9	-	-	-
D15-P10	108	1,900	-
D15-P11	-	-	-
D15-P12	1,052	6,100	-
D15-P13	105	6,200	56
D15-P14	-	-	-
D15-P15	-	-	-
D15-P16	48	-	-
D15-P17	-	-	-
D15-P18	32	4,800	-
D15-P19	882	24,500	-
D15-P20	-	-	-
D15-P21	-	-	-
D15-P22	-	-	-
D15-P23	78	-	-
D15-P24	103	-	-
D15-P25	-	-	-
D15-P26	572	6,700	-
D15-P27	274	7,505	68

TABLE 8

RABBIT AND GUINEA PIG ANTIBODY RESPONSES TO D15 PEPTIDES

Immunogen	Peptide-specific ELISAs	
	Reactive Titer ¹	
	Rabbit ²	Guinea Pig ³
D15-P1	102,400	819,200
D15-P2	204,800	1,637,400
D15-P3	51,200	1,637,400
D15-P4	204,800	819,200
D15-P5	51,200	1,637,400
D15-P6	51,200	409,600
D15-P7	204,800	819,200
D15-P8	51,200	409,600
D15-P9	102,400	409,600
D15-P10	102,400	819,200
D15-P11	51,200	819,200
D15-P12	102,400	204,800
D15-P13	NT ⁴	204,800
D15-P14	NT	409,600
D15-P15	NT	204,800
D15-P16	NT	819,200
D15-P17	NT	204,800
D15-P18	NT	312,500
D15-P19	NT	312,500
D15-P20	NT	62,500
D15-P21	NT	62,500
D15-P22	NT	12,500
D15-P23	NT	1,562,500
D15-P24	NT	312,500
D15-P25	NT	62,500

TABLE 9

RABBIT IgG ANTIBODY RESPONSE TO D15-PRP CONJUGATE

Rabbit ¹ #	Reactive Titer Against ²			
	PRP		rD15	
	2 doses	3 doses	2 doses	3 doses
489-1	1,600	3,200	1,600	6,400
490-1	1,600	1,600	6,400	25,600

¹ Rabbits were immunized intramuscularly with rD15-PRP conjugates (5 to 50 µg PRP equivalent) mixed with 3 mg ALPO₄ per mL, followed by two booster doses (half amount of the same immunogen) at 2 week intervals.

² Reactive titres is based on PRP specific and D-15 specific ELISAs.

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8. The recombinant vector of claim 6 wherein said DNA segment encodes a polypeptide of at least 6 residues.
9. The recombinant vector of claim 8 wherein said polypeptide is selected from those shown in Table 2.
10. The recombinant vector of claim 6, 7, 8 or 9 wherein said DNA segment consists of no more than the coding sequence for said D15 outer membrane protein.
11. The recombinant vector of claim 10, wherein the DNA segment further comprises a nucleic acid sequence encoding a leader sequence for export of said gene product from said host.
12. A purified and isolated protein encoded by the DNA fragment contained in the recombinant vector of claim 10 or 11.
13. A purified and isolated D15 outer membrane protein, or a portion thereof.
14. The protein of claim 13 wherein the D15 outer membrane protein is a Haemophilus D15 outer membrane protein.
15. The protein of claim 14 wherein the D15 outer membrane protein is a Haemophilus influenzae D15 outer membrane protein.
16. The protein of claim 15 wherein the Haemophilus influenzae is a type b Haemophilus influenzae strain.
17. The protein of claim 16 wherein the Haemophilus influenzae type b strain is selected from Ca, MinnA and Eagan strains.
18. The protein of claim 15 wherein the Haemophilus influenzae is a non-typeable Haemophilus influenzae strain.
19. The protein of claim 18 wherein the non-typeable Haemophilus influenzae strain is selected from PAK12085 and SB33 strains.
20. A synthetic peptide containing an amino acid sequence corresponding to the amino acid sequence of the protein or portion thereof claimed in any one of claims

30. The chimeric molecule of claim 29 wherein said another polypeptide or protein comprises a P1, P2 or P6 outer membrane protein of H. influenzae.

31. The chimeric molecule of claim 28 wherein said polysaccharide comprises a PRP molecule from H. influenzae.

FIG. 1A.

H. influenzae b Ca strain D15 sequence

-35
Hind III

GATTACGCCAAGCCTTAACGGTGTTTGGCAATTATTAAATGATTTTACGCTCTAATAATTAT
 10 20 30 40 50 60

RBS MET LYS LYS LEU LEU ILE ALA SER LEU LEU PHE GLY THR THR T
 AATAGGATACAAATCGATGAATAAACTTCTAATCGCAAGTTTATTATTCGGTACGACAACGA
 70 80 90 100 110 120

start truncated GST/D15 →

HR VAL PHE ALA ALA PRO PHE VAL ALA LYS ASP ILE ARG VAL ASP GLY VAL GLN THR ASP L
 CTGTGTTTGCCGCACCTTTTGTGGCAAAAGATAATTCCGTGTGGATGGTGTTCAGGTGACT
 130 140 150 160 170 180

EU GLU GIN GIN ILE ARG ALA SER LEU PRO VAL ARG ALA GLY GIN ARG VAL THR ASP ASN A
 TAGAACAAACAATCCGAGCAAGTTTACCTGTTCGTCGCCGTCAGCGTGTGACTGACAATG
 190 200 210 220 230 240

spurious thrombin site

SP VAL ALA ASN ILE VAL ARG | SER LEU PHE VAL SER GLY ARG PHE ASP VAL LYS ALA H
 ATGTGGCTAATAATTGTCCGCCTCTTTATTTCGTAAGTGGTCGATTCCGATGATGTGAAGCGC
 250 260 270 280 290 300

IS GIN GLU GLY ASP VAL LEU VAL VAL SER VAL ALA LYS SER ILE SER ASP VAL L
 ATCAAGAAAGGCGATGTGCTTGTGTTAGCGTTGTGGCTAAATCGATCATTTTCAGATGTTA
 310 320 330 340 350 360

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FIG.1A.(CONTINUED)

YS ILE LYS GLY ASN SER VAL ILE PRO THR GLU ALA LEU LYS GLN ASN LEU ASP ALA ASN G
 AATCAAGGTAACCTCTGTATATCCCACTGAAGCACTTAACAACCTTAGATGCTAACG 410 420
 370 380 390 400
 LY PHE LYS VAL GLY ASP VAL LEU ILE ARG GLU LYS LEU ASN GLU PHE ALA LYS SER VAL L
 GTTTAAAGTTGGCGATGTTTATAATTCGAGAGAAAATAAATGAATTGCCCCAAAAGTGTTAA 460 470 480
 430 440 450
 YS GLU HIS TYR ALA SER VAL GLY ARG TYR ASN ALA THR VAL GLU PRO ILE VAL ASN THR L
 AGAGCACTATGCAAGTGTAAGGTCGCTATAACGCAACAGTTGAACCTATTGTCAATACGC 510 520 530 540
 490 500
 EU PRO ASN ASN ARG ALA GLU ILE LEU ILE GIN ILE ASN GLU ASP ASP LYS ALA LYS LEU A
 TACCAAATAATCGCGCTGAAATTTTAATTCAAAATCAATGAAGATGATAAAGCAAAATTGG 550 560 570 580 590 600
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 LA SER LEU THR PHE LYS GLY ASN GLU SER VAL SER SER THR LEU GIN GLU GIN MET G
 CATCACTTAACCTTTCAAGGGGAACGAATCTGTAGTAGCACTACATTACAAGAACAAATGG 610 620 630 640 650 660
 LU LEU GIN PRO ASP SER TRP TRP LYS LEU TRP GLY ASN LYS PHE GLU GLY ALA GIN PHE G
 AATTACAACCTGATCTCTGGTGGAATAATTATGGGGAAATAAATTTGAAGGTGGCGCAATTCCG 670 680 690 700 710 720
 end truncated GST/DI5
 LU LYS _ASP LEU GIN SER ILE ARG ASP TYR TYR LEU ASN ASN GLY TYR ALA LYS ALA GIN I
 AGAAAGATTTCAGCTCAATTTCGGTGATTATTATTAAATAATGGCTATGCCCCAAAGCACAAA 730 740 750 760 770 780

FIG.1A.(CONTINUED)

LE THR LYS THR ASP VAL GIN LEU ASN ASP GLU LYS THR LYS VAL ASN VAL THR ILE ASP V
 T T A C T A A A C G G A T G T T C A G C T A A A T G A T G A A A A C A A A A G T T A A T G T A A C C A T T G A T G
 790 800 810 820 830 840

AL ASN GLU GLY LEU GIN TYR ASP LEU ARG SER ALA ARG ILE ILE GLY ASN LEU GLY GLY M
 T A A T G A A G G T T T A C A G T A T G A C C T T C G T A G T G C A C G C A T T A T A G G T A A T C T G G G A G G T A
 850 860 870 880 890 900

ET SER ALA GLU LEU GLU PRO LEU LEU SER ALA LEU HIS LEU ASN ASP THR PHE ARG ARG S
 T G T C T G C C G A G C T T G A A C C T T T A C T T T C A G C A T T A C A T T T A A A T G A T A C T T C C G C C G T A
 910 920 930 940 950 960

ER ASP ILE ALA ASP VAL GLU ASN ALA ILE LYS ALA LYS LEU GLY GLU ARG GLY TYR GLY S
 G T G A T A T T G C A G A T G T A G A A A A T G C A A T T A A A G C A A A A C T T G G A G A A C G C G G T T A C G G T A
 970 980 990 1000 1010 1020

ER ALA THR VAL ASN SER VAL PRO ASP PHE ASP ALA ASN LYS THR LEU ALA ILE THR L
 G C G C A C C G G T A A A T T C A G T A C C T G A T T T T G A T G A T G C A A A T A A A C A T T A G C G A T A A C C C
 1030 1040 1050 1060 1070 1080

EU VAL VAL ASP ALA GLY ARG ARG LEU THR VAL ARG GIN LEU ARG PHE GLU GLY ASN THR V
 T T G T T G T G A T G C T G G A C G A C G T T A A C T G T T C G C C A A C T T C G C T T T G A A G G A A T A C C G
 1090 1100 1110 1120 1130 1140

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FIG.1A.(CONTINUED)

AL SER ALA ASP SER THR LEU ARG GLN GLU MET ARG GLN GLN GLY THR TRP TYR ASN S
 TTTCTGCTGATAGCACTTTACGTCAGGAATAAGCCCAACAAGAGGAACCTTGGTATAATT 1150
 1160 1170 1180 1190 1200
 ER GLN LEU VAL GLU LEU GLY LYS ILE ARG LEU ASP ARG THR GLY PHE PHE GLU THR VAL G
 CACAATTAGTTGAGTTAGGAAATAATTCGCTTAGATCGTACAGGTTTCTTCCGAAACAGTCCG 1210
 1220 1230 1240 1250 1260
 LU ASN ARG ILE ASP PRO ILE ASN GLY SER ASN ASP GLU VAL ASP VAL TYR LYS VAL L
 AAAACCGAATTGATCCCTATCAATGGGTAGTAATGATGAAGTGGAATGTCGTATATAAAGTCA 1270
 1280 1290 1300 1310 1320
 YS GLU ARG ASN THR GLY SER ILE ASN PHE GLY ILE GLY TYR GLU SER GLY ILE S
 AAGAACGGTAACACGGGGTAGTATCAACTTTGGGTATTGGTTACGGGTACAGAGAGTGGTATTA 1330
 1340 1350 1360 1370 1380
 ER TYR GLN ALA SER VAL LYS GLN ASP ASN PHE LEU GLY THR GLY ALA VAL SER ILE A
 GTTATCAAGCAAGTGTTAAACAAGATATAATTCTTGGGAACAGGGCGGCAGTAGTATAG 1390
 1400 1410 1420 1430 1440
 LA GLY THR LYS ASN ASP TYR GLY THR SER VAL ASN LEU GLY TYR THR GLU PRO TYR PHE T
 CTGGTACCGAAATAATGATTTATGGGTACGAGTGTCATAATTGGGTTATACCGAGCCCCATTATTA 1450
 1460 1470 1480 1490 1500
 HR LYS ASP GLY VAL SER LEU GLY GLY ASN VAL PHE PHE GLU ASN TYR ASP ASN SER LYS S
 CTAAGATGGGTGTAAGTCTTTGGTGGAAATGTTTCTTTTGAAACACTACGATATACTCTAAAAA 1510
 1520 1530 1540 1550 1560

FIG.1A.(CONTINUED)

ER ASP THR SER SER ASN TYR LYS ARG THR THR THR GLY SER ASN VAL THR LEU GLY PHE P
 GTGATACATCCTCTAATACTATAAGCGGTACGACTTACGGAGTAATGTTACTTTAGGTTTCC
 1570 1580 1590 1600 1610 1620

 RO VAL ASN GLU ASN ASN SER TYR TYR VAL GLY LEU GLY HIS THR THR TYR ASN LYS ILE SER A
 CTGTAAATGAATAATACTCTATATGTAGGATTAGGTCATACCTATAATAAATTAGTA
 1630 1640 1650 1660 1670 1680

 SN PHE ALA LEU GLU TYR ASN ARG ASN LEU TYR ILE GIN SER MET LYS PHE LYS GLY ASN G
 ACTTTGCTCTAGAAATATAACCGTAATTATATATTCATAATCAATGAAATTTAAAGGTAATG
 1690 1700 1710 1720 1730 1740

 LY ILE LYS THR ASN ASP PHE ASP PHE SER PHE GLY TRP ASN TYR ASN SER LEU ASN ARG G
 GCATTAAACAATAAGACTTTTGATTTTCTTTTGGTGGAACTATAACAGCCTTAATAGAG
 1750 1760 1770 1780 1790 1800

 LY TYR PHE PRO THR LYS GLY VAL LYS ALA SER LEU GLY GLY ARG VAL THR ILE PRO GLY S
 GCTATTCCCAACTAAAGGGGTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTT
 1810 1820 1830 1840 1850 1860

 ER ASP ASN LYS TYR TYR LYS LEU SER ALA ASP VAL GIN GLY GLY PHE TYR PRO LEU ASP ARG A
 CTGATAACAAATACTACAAACTAAGTGCAGATGTACAGGGTTTCTACCCATTAGACAGAG
 1870 1880 1890 1900 1910 1920

 SP HIS LEU TRP VAL VAL SER ALA LYS ALA SER ALA GLY TYR ALA ASN GLY PHE GLY ASN L
 ATCACCCTCTGGGTTGTATCTGCAAAAGCATCTGCAGGATATGCAATGGTTTGGAAACA
 1930 1940 1950 1960 1970 1980

FIG. 1A.(CONTINUED)

YS ARG LEU PRO PHE TYR GIN THR TYR THR ALA GLY GLY ILE GLY SER LEU ARG GLY PHE A
 AGCGTTTACCGTTCCTATCAAACTTATACAGCGGTGGCATCGGTTTCATTACGTGGTTTGTG 2000 2010 2020 2030 2040
 1990
 LA TYR GLY SER ILE GLY PRO ASN ALA ILE TYR ALA GLU TYR GLY ASN GLY SER GLY THR G
 CTTATGGTAGTATTGGACCTAACGCAATTATATGCCGGAATATGGTAATGGTAGTGGTACTG 2050 2060 2070 2080 2090 2100
 2050
 LY THR PHE LYS ILE SER SER ASP VAL ILE GLY GLY ASN ALA ILE ALA THR ALA SER A
 GTACTTTTAAAGAAGATAAGTTCTGATGTGATTGGTGAATGCAATCGCTACAGCTAGCG 2110 2120 2130 2140 2150 2160
 2110
 LA GLU LEU ILE VAL PRO THR PRO PHE VAL SER ASP LYS SER GIN ASN THR VAL ARG THR S
 CAGAGTTAATTGTGCCAACTCCATTGTGTGAGCGGATAAGAGCCAAATAACGGTCCGAACCT 2170 2180 2190 2200 2210 2220
 2170
 ER LEU PHE VAL ASP ALA ALA SER VAL TRP ASN THR LYS TRP LYS SER ASP LYS ASN GLY L
 CCTTATTGTGTTGATGCGGCAAGTGTTTGGAACTACTAAATGGAAATCAGATAAAATGGAT 2230 2240 2250 2260 2270 2280
 2230
 EU GLU SER ASP VAL LEU LYS ARG LEU PRO ASP TYR GLY LYS SER SER ARG ILE ARG ALA S
 TAGAGCGGATGTATTAAAGAAGATTGCCCTGATTATGGCAAAATCAAGCCGTTATTCCGGCCT 2290 2300 2310 2320 2330 2340
 2290

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FIG.1A.(CONTINUED)

ER THR GLY VAL GLY PHE GLN TRP GLN SER PRO ILE GLY PRO LEU VAL PHE SER TYR ALA L
 CTACAGGTGTCGGATTCCAAATGGCAATCTCCTATTGGGGCCATTGGTATTCTCTTATGCCCA 2350
 2360 2370 2380 2390 2400
 YS PRO ILE LYS LYS TYR GLU ASN ASP VAL GLU GLN PHE GIN PHE SER ILE GLY GLY S
 AACCAATTAAATAATATGAAATGATGATGTCGAACAGTTCCAAATTAGTATTGGAGGTT 2410
 2420 2430 2440 2450 2460
 ER PHE *** ***
 CTTTCTAATAAATTGAACCTTTTCTTCTCATCAGAACTCAAAACAACGTTCTCTGCCTAA 2470
 2480 2490 2500 2510 2520
 TTTAAATTGGGCAGAGAAATAATTAAACCCATCTTAATTAAAGGATATTATCAAATGAA 2530
 2540 2550 2560 2570 2580
 AACATCGCAAAAGTAACCGCACCTTGCTTTAGGTATTGCACTTGCTTCAGGCTATGCTTC 2590
 2600 2610 2620 2630 2640
 CGCTGAAGAAAAAATTGCTTTTCAATTAAATGCAGGTATATTTTCAACATCACCCAGATCGC 2650
 2660 2670 2680 2690 2700
 CAAGCGGTAGCAGATAAACTTGATGCTGAAATTTAAACCCTGTAGCTGAGAAATTAGCAGCA 2710
 2720 2730 2740 2750 2760

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FIG.1A.(CONTINUED)

AGCAAAAGAGTTGATGAATAAATTGCTGCTGCTCGTAAATAAGTAGAAGCAAAAGTT
2770 2780 2790 2800 2810 2820

GCGGCTTTAGAAAAGATGCCACCTCGCTTACGTCAAGCTGATATTCAAACGCGCAACAG
2830 2840 2850 2860 2870 2880

GAGATTAAATTAGGTGCGGCTGAAGATGCTGAATTACAATAATTGCAAGAACAA
2890 2900 2910 2920 2930 2940

GATAAAAAA

FIG.1B.

DS-712-2-1 DNA, Egan D15 sequence
IS THE SEQUENCE BEING TRANSLATED

ACAGGACAGCCTTCCCTTTTAACCTTGAAATAATTAGGGAATACTTCCTGGCGATTG 60
10 20 30 40 50

TCA TTAAATAATTAAAGTGGGCCCAATTCTATTGCAAAAGGTGCTGGCCCCATCAGCAAAAT 120
70 80 90 100 110

ATTGGATTGGTGTAATTTTAAAGTTTATGGCACTGATTAGTGTAATAATTAGGGATTATG 180
130 140 150 160 170 180 9/82

AATTTATTCCATTACCAAGTATTAGATGGCGGGTCATTAGTTTTTTTAAACAATGGAAGCT 240
190 200 210 220 230 240

GTTAAAGGAAACCTGTTTCTGAGCGGGTGCAAGCATCTGTTATCGAATTGGCGCAGCA 300
250 260 270 280 290 300

CTGTTAATAAGCTTAACGGTGTTTGCAATTATTTAATGATTTTACGCTATAATTATA 360
310 320 330 340 350 360

FIG.1B.(CONTINUED)

MET LYS LYS LEU LEU ILE ALA SER LEU LEU PHE GLY THR THR THR
 TAGGATACAAATCGATGAATAAAGTCTAATCGCAAGTTATTTATTCGGTACGACACGAC
 370 380 390 400 410 420
 VAL PHE ALA ALA PRO PHE VAL ALA LYS ASP ILE ARG VAL ASP GLY VAL GIN GLY ASP LEU
 TGTGTTTGGCCGACCTTTTGTGGCAAAAGATATTTCGTGTGGATGGTGTTCAGGTTCAAGGTGACTT
 430 440 450 460 470 480
 GLU GIN GIN ILE ARG ALA SER LEU PRO VAL ARG ALA GLY GIN ARG VAL THR ASP ASN ASP
 AGAACAAATAATCCGAGCAAGTTTACCTGTTCTGTCGTCGCGGTCAAGCGTGTGACTGACAAATGA
 490 500 510 520 530 540
 VAL ALA ASN ILE VAL ARG SER LEU PHE VAL SER GLY ARG PHE ASP VAL VAL LYS ALA HIS
 TGTGGCTAATAATTGTCGCTCTTTATTTCGTAAAGTGGTCCGATTCGATGATGTGAAAGCGCA
 550 560 570 580 590 600
 GIN GLU GLY ASP VAL LEU VAL VAL SER VAL VAL ALA LYS SER ILE ILE SER ASP VAL LYS
 TCAAGAGCGGATGTGCTTGTGTTAGCGTTGTGGCTTAAATCGATCATTTTCAGATGTTAA
 610 620 630 640 650 660
 ILE LYS GLY ASN SER VAL ILE PRO THR GLU ALA LEU LYS GIN ASN LEU ASP ALA ASN GLY
 AATCAAAGGTAACTCTGTATTTCCTCACTGAAGCACTTAAACAAACTTAGATGCTAACGG
 670 680 690 700 710 720

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FIG. 1B.(CONTINUED)

PHE LYS VAL GLY ASP VAL LEU ILE ARG GLU LYS LEU ASN GLU PHE ALA LYS SER VAL LYS
 GTT TAA AGT TGG CGA TGT TTT AAT TCG AGA A A A A T T A A A T G A A T T T G C C A A A A G T G T A A A
 730 740 750 760 770 780

GLU HIS TYR ALA SER VAL GLY ARG TYR ASN ALA THR VAL GLU PRO ILE VAL ASN THR LEU
 A G A G C A C T A T G C A A G T G T A G G T C G C T A T A A C G C A A C A G T T G A A C C T A T T G T C A A T A C G C T
 790 800 810 820 830 840

PRO ASN ASN ARG ALA GLU ILE LEU ILE GIN ILE ASN GLU ASP ASP LYS ALA LYS LEU ALA
 A C C A A A T A A T C G C G C T G A A A T T T A A T T C A A A T C A A T G A A G A T G A T A A A G C C A A A A T T G G C
 850 860 870 880 890 900

SER LEU THR PHE LYS GLY ASN GLU SER VAL SER SER THR LEU GIN GLU MET GLU
 A T C A T T A A C T T T C A A G G G A A C G A A T C T G T T A G T A G C A G T A C A T T A C A A G A C A A A T G G A
 910 920 930 940 950 960

LEU GIN PRO ASP SER TRP TRP LYS LEU TRP GLY ASN LYS PHE GLU GLY ALA GIN PHE GLU
 A T T A C A A C C T G A T T C T T G G T G G A A A T T A T G G G G A A A T A A A T T T G A A G G T G C G C A A T T C G A
 970 980 990 1000 1010 1020

LYS ASP LEU GIN SER ILE ARG ASP TYR TYR LEU ASN ASN GLY TYR ALA LYS ALA GIN ILE
 G A A A G A T T T G C A G T C A A T T C G T G A T T A T T A T T A A A T A A T G G C T A T G C C A A A G C A C A A A T
 1030 1040 1050 1060 1070 1080

THR LYS THR ASP VAL GIN LEU ASN ASP GLU LYS THR LYS VAL ASN VAL THR ILE ASP VAL
 T A C T A A A A C G G A T G T T C A G C T A A A T G A T G A A A A A C A A A A A G T T A A T G T A A C C A T T G A T G T
 1090 1100 1110 1120 1130 1140

FIG.1B.(CONTINUED)

ASN GLU GLY LEU GLN TYR ASP LEU ARG SER ALA ARG ILE ILE GLY ASN LEU GLY GLY MET
 AAATGAAGGTTTACAGTATGACCTTCGTAGTGCACGCATTATAGGTAATCTGGGAGGTAT 1150
 1160 1170 1180 1190 1200
 SER ALA GLU LEU GLU PRO LEU LEU SER ALA LEU HIS LEU ASN ASP THR PHE ARG ARG SER
 GTCTGCCGAGCTTGAAACCTTTACTTTTCAGCATTAACAATTAAATGATACTTCCGCCGTAG 1210
 1220 1230 1240 1250 1260
 ASP ILE ALA ASP VAL GLU ASN ALA ILE LYS ALA LYS LEU GLY GLU ARG GLY TYR GLY SER
 TGATATTGCAGATGTAGAAATGCAATTAAAGCAAACCTTGGAGAACGCCGTTACGGTAG 1270
 1280 1290 1300 1310 1320
 ALA THR VAL ASN SER VAL PRO ASP PHE ASP ALA ASN LYS THR LEU ALA ILE THR LEU
 CGCAACGGTAATTCAGTACCTGATTTTGATGATGCAATAAACAATTAGCGATAACCCCT 1330
 1340 1350 1360 1370 1380
 VAL VAL ASP ALA GLY ARG ARG LEU THR VAL ARG GIN LEU ARG PHE GLU GLY ASN THR VAL
 TGTGTGTGATGCTGGACGACGTTTAACCTGTTTCGCCCAACTTCGCTTTGAAGGAATAACCGT 1390
 1400 1410 1420 1430 1440
 SER ALA ASP SER THR LEU ARG GIN GIN GLU MET ARG GIN GIN GLY THR TRP TYR ASN SER
 TTCTGCTGATAGCACTTTACGTCAGGAAATGCCCAACAAGAACCTTGGTATAATTC 1450
 1460 1470 1480 1490 1500

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FIG.1B.(CONTINUED)

GLN LEU VAL GLU LEU GLY LYS ILE ARG LEU ASP ARG THR GLY PHE PHE GLU THR VAL GLU
 ACAATTAGTTGAGTTAGGAAAATTTCGCTTAGATCGTACAGGTTTCTTCGAAACAGTCGA 1550
 1510 1520 1530 1540 1550 1560

ASN ARG ILE ASP PRO ILE ASN GLY SER ASN ASP GLU VAL ASP VAL TYR LYS VAL LYS
 AACCGAATTGATCCTATCAATGGTAGTAATGATGAAGTGGAATGTCGTATATAAAGTCAA 1610
 1570 1580 1590 1600 1610 1620

GLU ARG ASN THR GLY SER ILE ASN PHE GLY ILE GLY TYR GLY THR GLU SER GLY ILE SER
 AGAACGTAAACACGGGTAGTATCAACTTTGGGTATTGGTTACGGTACAGAGAGTGGTATTAG 1670
 1630 1640 1650 1660 1670 1680

TYR GLN ALA SER VAL LYS GIN ASP ASN PHE LEU GLY THR GLY ALA VAL SER ILE ALA
 TTATCAAGCAAGTGTTAAACAAGATAATTCTTTGGGAACAGGGCGGCAGTAAGTATAGC 1730
 1690 1700 1710 1720 1730 1740

GLY THR LYS ASN ASP THR GLY THR SER VAL ASN LEU GLY TYR THR GLU PRO TYR PHE THR
 TGGTACGAAAATGATTATGGGTACGAGTGTCAAATTGGGTTATACCGAGCCCTATTTTAC 1790
 1750 1760 1770 1780 1790 1800

LYS ASP GLY VAL SER LEU GLY GLY ASN VAL PHE PHE GLU ASN TYR ASP ASN SER LYS SER
 TAAAGATGGTGTAAGTCTTGGTGGAATAATGTTTCTTTTGAAACACTACGATACTCTAAAG 1850
 1810 1820 1830 1840 1850 1860

FIG.1B.(CONTINUED)

ASP THR SER SER ASN TYR LYS ARG THR THR THR GLY SER ASN VAL THR LEU GLY PHE PRO
 TGATACATCCCTCTAATAAGCGTACGACTTACGGAAAGTAATGTTACTTTAGGTTTCCC 1880 1900 1910 1920

VAL ASN GLU ASN ASN SER TYR TYR VAL GLY LEU GLY HIS THR TYR ASN LYS ILE SER ASN
 TGTAAATGAATAAATACTCCTATTATGTAGGATTAGGTCAATACCTATAATAAATTAGTAA 1930 1940 1950 1960 1970 1980

PHE ALA LEU GLU TYR ASN ARG ASN LEU TYR ILE GLN SER MET LYS PHE LYS GLY ASN GLY
 CTTTGCTCTAGAAATATAACCGTAATTATATATCAATCAATGAAATTTAAAGGTAATGG 2000 2010 2020 2030 2040

ILE LYS THR ASN ASP PHE ASP PHE SER PHE GLY TRP ASN TYR ASN SER LEU ASN ARG GLY
 CATTAACAATAAGACTTTTGATTTTCTTTTGGTGGAACTATAACAGCCCTTAATAGAGG 2050 2060 2070 2080 2090 2100

TYR PHE PRO THR LYS GLY VAL LYS ALA SER PHE SER PHE GLY LEU GLY ARG VAL THR ILE PRO GLY SER
 CTATTTCCCAACTAAAGGGGTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTTTC 2110 2120 2130 2140 2150 2160

ASP ASN LYS TYR TYR LYS LEU SER ALA ASP VAL GIN GLY PHE TYR PRO LEU ASP ARG ASP
 TGATAACAATACTACAAACTAAGTGCAGATGTACAGGGTTCTTCTACCCATTAGACAGAGA 2170 2180 2190 2200 2210 2220

HIS LEU TRP VAL VAL SER ALA LYS ALA SER ALA GLY TYR ALA ASN GLY PHE GLY ASN LYS
 TCACCTCTGGGTTGTATCTGCAAAAGCATCTGCAGGATATGCAAATGGTTTGGGAAACAA 2230 2240 2250 2260 2270 2280

FIG.1B.(CONTINUED)

ARG LEU PRO PHE TYR GLN THR TYR THR ALA GLY GLY ILE GLY SER LEU ARG GLY PHE ALA
 GCGTTTACCGTTCTATCAAACTTATACAGCGGGTGGCAATCGGTTTCATTACGTGGTTTGGC 2290 2300 2310 2320 2330 2340

TYR GLY SER ILE GLY PRO ASN ALA ILE TYR ALA GLU TYR GLY ASN GLY SER GLY THR GLY
 TTA TGGTAGTATTGGACCTAACGCAATTATATGCCGAATATGGTAATGGTAGTGGTACTGG 2350 2360 2370 2380 2390 2400

THR PHE LYS LYS ILE SER SER ASP VAL ILE GLY GLY ASN ALA ILE ALA THR ALA SER ALA
 TACTTTTAAAGAAAGATAAGTTCCTGATGTGATTGGTGGTAATGCATAATCGCTACAGCTAGCGC 2410 2420 2430 2440 2450 2460

GLU LEU ILE VAL PRO THR PRO PHE VAL SER ASP LYS SER GLN ASN THR VAL ARG THR SER
 AGAGTTAATTGTGCCCAACTCCATTGTGAGCGGATAAGAGCCCAAATAACGGTCCGAACCTC 2470 2480 2490 2500 2510 2520

LEU PHE VAL ASP ALA ALA SER VAL TRP ASN THR LYS TRP LYS SER ASP LYS ASN GLY LEU
 CTTATTGTGTTGATGCGGCAAGTGTTTGGGAATACTAAATGGAAATCAGATAAATAATGGATT 2530 2540 2550 2560 2570 2580

GLU SER ASP VAL LEU LYS ARG LEU PRO ASP TYR GLY LYS SER SER ARG ILE ARG ALA SER
 AGAGAGCGGATGTATTAAAGATTTGCCCTGATTATGGCAAAATCAAGCCGTAATTCGCGCCCTC 2590 2600 2610 2620 2630 2640

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FIG.1B.(CONTINUED)

THR GLY VAL GLY PHE GLN TRP GLN SER PRO ILE GLY PRO LEU VAL PHE SER TYR ALA LYS
TACAGGTGTCGGAATCCCAATGGCAATCTCCTATTGGGCCCATTTGGTATTCTCTTATGCCCAA 2690 2700

PRO ILE LYS LYS TYR GLU ASN ASP VAL GLU GLN PHE SER ILE GLY SER
ACCAATTAAAAATATGAAAAATGATGATGTCGAAACAGTTCCAATTAGTATTGGAGGTTTC 2740 2750 2760

PHE *** ***

TTTCTAATAAATTGAACTTTTCTTCTCATCAGAACTCAAAACAACGTTCTCTGCCCTAAT 2770 2780 2790 2800 2810 2820

TTAATTGGGCAGAGAAAATATTAAACCCCATCTTAAATTAGGATATTATCAAAATGAAA 2830 2840 2850 2860 2870 2880

AACATCGCAAAAGTAACCGCACTTGCTTTAGGTATTGCACCTTGCTTCAGGCTATGCTTCC 2890 2900 2910 2920 2930 2940

GCTGAAGAAAAAATTGCTTTTCATTAAATGCACCTATTTTTC AA 2950 2960 2970 2980

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FIG.1C.

DS-691-1-5 DNA, Mirm A D15 sequence
IS THE SEQUENCE BEING TRANSLATED

AATCACTTACTGGCGATTGTCTAATAATAATTAAAGTGGGCCAATTCTATTGCAAAAG
10 20 30 40 50 60

GTGCTGGCACATCAGCAAATATTGGATTGGGTGTATTTTAAAGTTTATGGCACTGATTA
70 80 90 100 110 120

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GTGTAAATTAGGGATTATGAATTATTTCCATTACCAGTATTAGATGGCGGTCATTAG
130 140 150 160 170 180

TTTTTTAAACAATGGGAAGCTGTAAAGGAAACCTGTTCTGAGCGGGTGCAAGCATCT
190 200 210 220 230 240

GTTATCGAATTGGCGCAGCACTGTTATTAAAGCTTAACGGTGTTTGCAATTATAATGATT
250 260 270 280 290 300

FIG.1C.(CONTINUED)

GLN ASN LEU ASP ALA ASN GLY PHE LYS VAL GLY ASP VAL LEU ILE ARG GLU LYS LEU ASN
 CAAACTTAGATGCTAACGGGTTTAAAGTTGGCGGATGTTTAAATTCGAGAAAAATTAAAT
 670 680 690 700 710 720

GLU PHE ALA LYS SER VAL LYS GLU HIS TYR ALA SER VAL GLY ARG TYR ASN ALA THR VAL
 GAATTTGCCCAAAGTGTAAGAGCAGCTATGCAAGTGTAAGTCGCTATAACGCAACAGTT
 730 740 750 760 770 780

GLU PRO ILE VAL ASN THR LEU PRO ASN ASN ARG ALA GLU ILE LEU ILE GIN ILE ASN GLU
 GAACCTATTGTCAATAACGCTACCAATAATCGCGCTGAAATTTTAATTCAAAATCAATGAA
 790 800 810 820 830 840

ASP ASP LYS ALA LYS LEU ALA SER LEU THR PHE LYS GLY ASN GLU SER VAL SER SER
 GATGATAAGCAAAATTGGCATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGT
 850 860 870 880 890 900

THR LEU GIN GLU GIN MET GLU LEU GIN PRO ASP SER TRP TRP LYS LEU TRP GLY ASN LYS
 ACATTACAAGAACAAATGGAAATTACAACCTGATTCCTTGGTGGGAAATTAATGGGGGAAATAAA
 910 920 930 940 950 960

PHE GLU GLY ALA GIN PHE GLU LYS ASP LEU GIN SER ILE ARG ASP TYR TYR LEU ASN ASN
 TTTGAAGGTGCGCAATTTCGAGAAAGATTTCAGTCAATTTCGTGATTATTATTAAATAAT
 970 980 990 1000 1010 1020

FIG.1C.(CONTINUED)

GLY TYR ALA LYS ALA GLN ILE THR LYS THR ASP VAL GIN LEU ASN ASP GLU LYS THR LYS
 GGCTATGCCCAAAGCACAAATTACTTAAACGGATGTTTCAGCTAAATGATGAATAAACAAAA
 1030 1040 1050 1060 1070 1080

VAL ASN VAL THR ILE ASP VAL ASN GLU GLY LEU GIN TYR ASP LEU ARG SER ALA ARG ILE
 GTTAATGTAACCATTCGATGTAAATGAAGGTTTACAGTATGACCTTCGGTAGTGACGCAATT
 1090 1100 1110 1120 1130 1140

ILE GLY ASN LEU GLY GLY MET SER ALA GLU LEU GLU PRO LEU LEU SER ALA LEU HIS LEU
 ATAGGTAATCTGGAGGTAATGCTCTGCCGAGCTTGAAACCTTTACTTTCAGCAATTACATTTA
 1150 1160 1170 1180 1190 1200

20/82

ASN ASP THR PHE ARG ARG SER ASP ILE ALA ASP VAL GLU ASN ALA ILE LYS ALA LYS LEU
 AATGATACTTCCGCCGCTAGTGATATTGCGAGATGTAGAAAATGCAATTAAAGCAAAACTT
 1210 1220 1230 1240 1250 1260

GLY GLU ARG GLY TYR GLY SER ALA THR VAL ASN SER VAL PRO ASP PHE ASP ALA ASN
 GGAGAACGGCGGTACGGTAGCGCAACGGTAAATTCAGTACCTGATTTTGATGATGCAAAAT
 1270 1280 1290 1300 1310 1320

LYS THR LEU ALA ILE THR LEU VAL VAL ASP ALA GLY ARG ARG LEU THR VAL ARG GLN LEU
 AAAACATTAGCGGATAACCCCTTGTGTTGATGCTGGACGCGTTTAACTGTTCCGCCAACTT
 1330 1340 1350 1360 1370 1380

FIG.1C.(CONTINUED)

ARG PHE GLU GLY ASN THR VAL SER ALA ASP SER THR LEU ARG GLN GLU MET ARG GLN GIN
 CGCTTTGAAGGAAATACCGTTTCTGCTGATAGCACTTTACGTCAGGAAATGCGCCAAACA
 1390 1400 1410 1420 1430 1440

GLU GLY THR TRP TYR ASN SER GLN LEU VAL GLU LEU GLY LYS ILE ARG LEU ASP ARG THR
 GAAGGAACCTTGGGTATTAATTCACAAATTAGTTGAGTTAGGAAATAATTCGCTTAGATCGTACA
 1450 1460 1470 1480 1490 1500

GLY PHE PHE GLU THR VAL GLU THR VAL GLU ASN ARG ILE ASP PRO ILE ASN GLY SER ASN ASP GLU VAL
 GGTTCCTTCGAAACAGTCGAAACCGAAATTGATCCCTATCAATGGTAGTAATGATGAAGTG
 1510 1520 1530 1540 1550 1560

ASP VAL VAL TYR LYS VAL LYS GLU ARG ASN THR GLY SER ILE ASN PHE GLY ILE GLY TYR
 GATGTCGTATATAAAGTCAAGAACGTAACACGGGTAGTATCAACTTTGGTATTGGTTAC
 1570 1580 1590 1600 1610 1620

GLY THR GLU SER GLY ILE SER TYR GLN ALA SER VAL LYS GIN ASP ASN PHE LEU GLY THR
 GTACAGAGAGTGGTATTAGTTATCAAGCAAGTGTTAAACAAGATAATTCTTGGGAACA
 1630 1640 1650 1660 1670 1680

GLY ALA ALA VAL SER ILE ALA GLY THR LYS ASN ASP TYR GLY THR SER VAL ASN LEU GLY
 GGGCGGCAGTAAGTATAGCTGGGTACGAAATAATGATTATGGTACGAGTGTCAAATTGGGT
 1690 1700 1710 1720 1730 1740

FIG.1C.(CONTINUED)

TYR THR GLU PRO TYR PHE THR LYS ASP GLY VAL SER LEU GLY GLY ASN VAL PHE PHE GLU
 TATACCGAGCCCTATTCTTACTAAAGATGGTGTAAGTCTTGGGTGGAAATGTTTCTTTGAA
 1750 1760 1770 1780 1790 1800

 ASN TYR ASP ASN SER LYS SER SER ASP THR SER SER SER ASN TYR LYS ARG THR THR TYR GLY SER
 AACTACGATAACTCTAAAGTGATACATCCCTCTAACTATAAGCGTACGACTTACGGAAGT
 1810 1820 1830 1840 1850 1860

 ASN VAL THR LEU GLY PHE PRO VAL ASN GLU ASN ASN SER TYR TYR VAL GLY LEU GLY HIS
 AATGTTACTTTAGGTTTCCCTGTAAATGAAAATAAATCCTATTTATGTAGGATTAGGTCAT
 1870 1880 1890 1900 1910 1920

 THR TYR ASN LYS ILE SER ASN PHE ALA LEU GLU TYR ASN ARG ASN LEU TYR ILE GLN SER
 ACCATAATAAATTAGTAACCTTTGCTCTAGAAATAAACCGTAATTATATATATCAATCA
 1930 1940 1950 1960 1970 1980

 MET LYS PHE LYS GLY ASN GLY ILE LYS THR ASN ASP PHE ASP PHE SER PHE GLY TRP ASN
 ATGAAATTTAAGGTAATGGCATTTAAACAATAAGACTTTTGATTTTCTTTTGGTTGGAAAC
 1990 2000 2010 2020 2030 2040

 TYR ASN SER LEU ASN ARG GLY TYR PHE PRO THR LYS GLY VAL LYS ALA SER LEU GLY GLY
 TATAACAGCCCTTAATAGAGGCTATTTCCTCAACTAAAGGGGTTAAAGCAAGCTCTTGGTGGA
 2050 2060 2070 2080 2090 2100

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FIG.1C.(CONTINUED)

ARG VAL THR ILE PRO GLY SER ASP ASN LYS TYR TYR LYS LEU SER ALA ASP VAL GLN GLY
 CGAGTTACTATTCCAGGTTCTGTGATAACAATACTACAACTAAGTGCAGATGTACAGGGT 2110 2120 2130 2140 2150 2160

 PHE TYR PRO LEU ASP ARG ASP HIS LEU TRP VAL VAL SER ALA LYS ALA SER ALA GLY TYR
 TTCTACCCATTAGACAGAGATCACCTCTGGGTTGTATCTGTGCAAAAGCATCTGCAGGATAT 2170 2180 2190 2200 2210 2220

 ALA ASN GLY PHE GLY ASN LYS ARG LEU PRO PHE TYR GLN THR TYR THR ALA GLY ILE
 GCAAAATGGTTTGGGAAACAAGCGTTTACCGTTTCTATCAAACTTATACAGCGGGTGGCATC 2230 2240 2250 2260 2270 2280

 GLY SER LEU ARG GLY PHE ALA TYR GLY SER ILE GLY PRO ASN ALA ILE TYR ALA GLU TYR
 GGTTCATTACGTGGTTTTCCTTATGGTAGTATTGGACCTAACGCAATTATGCCGAATAT 2290 2300 2310 2320 2330 2340

 GLY ASN GLY SER GLY THR GLY THR PHE LYS LYS ILE SER SER ASP VAL ILE GLY GLY ASN
 GGTAATGGTAGTGGTACTGGTACTTTTAAAGAAGATAAGTTCTGTGATTGGTGGTAAT 2350 2360 2370 2380 2390 2400

 ALA ILE ALA THR ALA SER ALA GLU LEU ILE VAL PRO THR PRO PHE VAL SER ASP LYS SER
 GCAATCGCTACAGCTAGCGCAGAGTTAATTGTGCCAACTCCATTGTGAGCGGATAAGAGC 2410 2420 2430 2440 2450 2460

23/82

FIG.1C.(CONTINUED)

GLN ASN THR VAL ARG THR SER LEU PHE VAL ASP ALA ALA SER VAL TRP ASN THR LYS TRP
 CAA AATACGGTCCGGAACCTCCTTATTGTTGATGCGGCAAGTGT TTGGAAATACTAAATGG
 2470 2480 2490 2500 2510 2520

 LYS SER ASP LYS ASN GLY LEU GLY SER LEU GLY LEU LYS ARG LEU PRO ASP TYR GLY LYS
 AAATCAGATAAAATGGATTAGAGAGCGGATGTTAA AAGATTGCCCTGATTATGGCAAA
 2530 2540 2550 2560 2570 2580

 SER SER ARG ILE ARG ALA SER THR GLY VAL GLY PHE GLN TRP GIN SER PRO ILE GLY PRO
 TCAAGCCGTATTCCGCCCTCTACAGGTGTCGGATTCCAAATGGCAATCTCCTATTGGGCCCA
 2590 2600 2610 2620 2630 2640

 LEU VAL PHE SER TYR ALA LYS PRO ILE LYS LYS TYR GLU ASN ASP ASP VAL GLU GIN PHE
 TTGGTATTCTCTTATGCCCAAACCAATTAA AATAATGAAATGATGATGTCGAACAGTTC
 2650 2660 2670 2680 2690 2700

 GLN PHE SER ILE GLY GLY SER PHE ***
 CAATTAGTATTGGAGGTTCTTCTTAATAAATTGAACTTTTCTTCTCATCAGAACTCAA
 2710 2720 2730 2740 2750 2760

24/82

FIG.1C.(CONTINUED)

AACAACGTTCTCTGCCCTAATTTAATTGGGGCAGAGAAATAATTAAACCCATCATTTAATA
2770 2780 2790 2800 2810 2820

AGGATATTTATCAAAATGAAAAACATCGCAAAAGTAACCGCACTTGCTTTAGGTAATTGCAC
2830 2840 2850 2860 2870 2880

TTGCTTCAGGCTATGGCTTCCGCTGAAGAAAAAATTGCTTTCATTAAATGCCGGGTATANTTT
2890 2900 2910 2920 2930 2940 25/82

TNCAAGGCNAAGG
2950

FIG.1D.

SB33 D15

IS THE SEQUENCE BEING TRANSLATED

GGCA TTGAAA AAACAGGACAGC TTTC CCTTTT AACCTTGAAA AATA TTAGGGA AATTACTT
10 20 30 40 50 60

ACTGGCGA TTGTCA TTAAATAA TTAAAGTGGGCCAATTCTAT TGCAAAAGGTGCTGGC
70 80 90 100 110 120

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GCATCAGCAA ATAATTGGATTGGTGTA TTTTAAAGTTTATGGCATTGATTAGTGTA AAT
130 140 150 160 170 180

TTAGGGATTATGAATTTATTTCCATTACCAGTATTAGATGGCGGTCATTTAGTTT TTTTA
190 200 210 220 230 240

ACAA TGGAAGCTGTAAAGGAAAACCTGTTTCTGAGCGGGTGCA AAGCATCTGTTATCGA
250 260 270 280 290 300

FIG.1D.(CONTINUED)

ATTGGCGCAGCACTGTTATTAAAGCTTAACGGGTGTTTGCAATTATTAAATGATTTTACGT 310 320 330 340 350 360

CTATAATTATATAGGATACAAATCGATGAAAAAACAATCTTAATCGCAAGTTTATTATTCCGG 370 380 390 400 410 420

THR THR THR THR VAL PHE ALA ALA PRO PHE VAL ALA LYS ASP ILE ARG VAL ASP GLY VAL 27/82
TACGACAACGACTGTGTTTGCCCGCACCTTTTGTGGCAAAAGATATTCTGTTGTGGATGGTGT 430 440 450 460 470 480

GLN GLY ASP LEU GLU GIN GIN ILE ARG ALA SER LEU PRO VAL ARG ALA GLY GIN ARG VAL
TCAAGGTGACTTAGAACAAACAATCCGAGCAAGTTTACCCTGTTCTGTCGCCGGTTCAGCGTGT 490 500 510 520 530 540

THR ASP ASN ASP VAL ALA ASN ILE VAL ARG SER LEU PHE VAL SER GLY ARG PHE ASP ASP
GACTGACAAATGATGTGGCTAATAATTGTCCGCCCTCTTTATTCTGTAAGTGGTCCGATTCGATGA 550 560 570 580 590 600

VAL LYS ALA HIS GIN GLU GLY ASP VAL LEU VAL VAL SER VAL VAL ALA LYS SER ILE ILE
TG TGAAAGCGCATCAAGAGGCGGATGTGCTTGTGTTAGCGTTGTGGCTAAATCGATCAT 610 620 630 640 650 660

FIG.1D.(CONTINUED)

SER ASP VAL LYS ILE LYS GLY ASN SER ILE ILE PRO PRO GLU ALA LEU LYS GLN ASN LEU
 TTCAGATGTTAAATCAAAAGGTAACCTCTATTATTCACCTGAAGCACTAAACAAACTT
 670 680 690 700 710 720

ASP ALA ASN GLY PHE LYS VAL GLY ASP ILE ILE LEU ILE ARG GLU LYS LEU ASN GLU PHE ALA
 AGATGCTAACGGGTTTAAAGTTGGCGATATTTTAATTCGAGAAATAATTAATGAATTGTC
 730 740 750 760 770 780

GLN SER VAL LYS GLU HIS TYR ALA SER VAL GLY ARG TYR ASN ALA THR VAL GLU PRO ILE
 CCAAAGTGTAAGAGCACTATGCAAGTGTAAGTTCGGCTATTAACGCCAACCGTTGAACCTAT
 790 800 810 820 830 840

VAL ASN THR LEU PRO ASN ASN ARG ALA GLU ILE ILE LEU ILE ASN GLU ASP ASP LYS
 TGTCAATACGCTACCAATAATCAGCGCTGAAATTTTAATTCAATCAATGAAGATGATAA
 850 860 870 880 890 900

ALA LYS LEU ALA SER LEU THR PHE LYS GLY ASN GLU SER VAL SER SER THR LEU GLN
 AGCCAAATTGGCATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGTACATTACA
 910 920 930 940 950 960

GLU GLN MET GLU LEU GLN PRO ASP SER TRP TRP LYS LEU TRP GLY ASN LYS PHE GLU GLY
 AGAACAAATGGAAATTACAACCTGATTCTTGGGTGGAAATTAATGGGGAATAAATTTGAAGG
 970 980 990 1000 1010 1020

FIG.1D.(CONTINUED)

ALA GIN PHE GLU LYS ASP LEU GIN ALA ILE ARG ASP TYR TYR LEU ASN ASN GLY TYR ALA
 TCGGCAATTTCGAGAAAGATTTCGACGGCAATTTCGTGATTATTTAAATAATGGCTATGCT
 1030 1040 1050 1060 1070 1080

LYS ALA GIN ILE THR LYS ALA ASP VAL GIN LEU ASN ASP GLU LYS THR LYS VAL ASN VAL
 CAAAGCACAAATCACTAAAGCGGATGTTTCAGCTAAATGATGAAAAACAAAGTTAATGT
 1090 1100 1110 1120 1130 1140

THR ILE ASP VAL ASN GLU GLY LEU GIN TYR ASP LEU ARG SER ALA ARG ILE ILE GLY ASN
 ACCATTGATGTAATAAGGTTTACAGTATGACCTTCGTAGTGACGCATTATAGGTAA
 1150 1160 1170 1180 1190 1200

LEU GLY GLY MET SER ALA GLU LEU GLU PRO LEU LEU SER ALA LEU HIS LEU ASN ASP THR
 TCTGGGAGGTATGTCCTGCGGAGCCTTGAAACCTTTACTTTTCAGCATTACATTAAATGATAC
 1210 1220 1230 1240 1250 1260

PHE ARG ARG SER ASP ILE ALA ASP VAL GLU ASN ALA ILE LYS ALA LYS LEU GLY GLU ARG
 TTCCGCCGTAGTGATATTTCGAGATGTAGAAATGCAATTAAAGCAAAACTTGGGGAACG
 1270 1280 1290 1300 1310 1320

GLY TYR GLY ASN THR THR VAL ASN SER VAL PRO ASP PHE ASP ALA ASN LYS THR LEU
 AGGTACGGTAACACAAACAGTAAATTCTGTACCTGATTTTGACGATGCAATAAACAATT
 1330 1340 1350 1360 1370 1380

29/82

FIG.1D.(CONTINUED)

ALA ILE THR PHE VAL VAL ASP ALA GLY ARG ARG LEU THR VAL HIS GLN LEU ARG PHE GLU
 1390 1400 1410 1420 1430 1440

ACCGATAACCTTTGTTGTTGATGCTGGACGACGTTTAACTGTTCAACCAACTTCGCTTTTGA

GLY ASN THR VAL SER ALA ASP SER THR LEU ARG ARG MET ARG GLN GLN GLU GLY THR
 1450 1460 1470 1480 1490 1500

AGGAAATACCGTTTCTGCTGATAGTACTTTACGTCAGGAATAATGCCCAACAAGAGGAAC

TRP TYR ASN SER GIN LEU VAL GLU LEU GLY LYS ILE ARG LEU ASP ARG THR GLY PHE
 1510 1520 1530 1540 1550 1560

TGGTATAATTCACAATTAGTTGAGTTAGGAATAATTCGCTTAGATCGTACAGGTTTCTT

GLU THR VAL GLU ASN ARG ILE ASP PRO ILE ASN GLY SER ASN ASP GLU VAL ASP VAL
 1570 1580 1590 1600 1610 1620

CGAAACAGTTGAAACCCGAATTGATCCCTATCAATGGGTAGCAATGATGAAGTGGAATGTCGT

TYR LYS VAL LYS GLU ARG ASN THR GLY SER ILE ASN PHE GLY ILE GLY TYR GLY THR GLU
 1630 1640 1650 1660 1670 1680

ATATAAGTCAAGAACGTAACACGGGTAGTATCAACTTTGGTATTGGTTACGGTACAGA

SER GLY ILE SER TYR GIN ALA SER VAL LYS GIN ASP ASN PHE LEU GLY THR GLY ALA ALA
 1690 1700 1710 1720 1730 1740

GAGTGGTATTAGTTATCAAGCAAGTGTCACAACAAGATAATTCTTTGGGAACAGGGGGCGGC

FIG.1D.(CONTINUED)

VAL SER ILE ALA GLY THR LYS ASN ASP TYR GLY THR SER VAL ASN LEU GLY TYR THR GLU
 AGTAAGTATAGCTGGTACGAAATAATGATTATGGTACGAGTGTCAATTGGGTTATACCGA
 1750 1760 1770 1780 1790 1800

PRO TYR PHE THR LYS ASP GLY VAL SER LEU GLY GLY PHE VAL PHE GLU ASN TYR ASP
 GCCCTATTCTACTAAAGATGGTGTAAGTCTTGGTGGAAATGTTTCTTTGAAACTACGA
 1810 1820 1830 1840 1850 1860

ASN SER LYS SER ASP THR SER SER SER ASN TYR LYS ARG THR THR TYR GLY SER ASN VAL THR
 TAACTCTAAAGTGATACATCCCTCTAATACTATAAGCGTACGACTTATGGAACTAATGTTAC
 1870 1880 1890 1900 1910 1920

LEU GLY PHE PRO VAL ASN GLU ASN ASN SER TYR TYR VAL GLY LEU GLY HIS THR TYR ASN
 TTTAGGTTTCCCTGTAAATGAATAAATACTCCTATTATGTAGGATTAGGCCATACCTATAA
 1930 1940 1950 1960 1970 1980

LYS ILE SER ASN PHE ALA LEU GLU TYR ASN ARG ASN LEU TYR ILE GLN SER MET LYS PHE
 TAAATTAGTAACCTTTGCTCTAGAAATATAACCGTAATTATATATCAATCAATGAAATT
 1990 2000 2010 2020 2030 2040

LYS GLY ASN GLY ILE LYS THR ASN ASP PHE ASP PHE GLY TRP ASN TYR ASN SER
 TAAAGGTAAATGGCATTAACAACAATAAGACTTTTGATTTTCTTTTGGTTGGAACTATAACAG
 2050 2060 2070 2080 2090 2100

FIG.1D.(CONTINUED)

LEU ASN ARG GLY TYR PHE PRO THR LYS GLY VAL LYS ALA SER LEU GLY ARG VAL THR
 CCTTAAATAGAGGCTATTTCCTCAACTAAAGGGGTTAAAGCAAGTCTTGGTGGACGAGTTAC
 2110 2120 2130 2140 2150 2160

ILE PRO GLY SER ASP ASN LYS TYR TYR LYS LEU SER ALA ASP VAL GLN GLY PHE TYR PRO
 AATTCAGGTTCTGATAACAAATACTACAAACTAAGTGCAGATGTACAGGGTTTCTTACCC
 2170 2180 2190 2200 2210 2220

LEU ASP ARG ASP HIS LEU TRP VAL VAL SER ALA LYS ALA SER ALA GLY TYR ALA ASN GLY
 ATTAGACAGAGATCACCTCTGGGTTGTATCTGCAAAAGCATCTGCAGGATATGCAAAATGG
 2230 2240 2250 2260 2270 2280

PHE GLY ASN LYS ARG LEU PRO PHE TYR GLN THR TYR THR ALA GLY GLY ILE GLY SER LEU
 TTTTGGAAACAAGCGTTTACCGTTCTATCAAACTTATACAGCGGGTGGCATTTGGTTCATT
 2290 2300 2310 2320 2330 2340

ARG GLY PHE ALA TYR GLY SER ILE GLY PRO ASN ALA ILE TYR GLN GLY GLN ASN ASN LYS
 ACGCGGTTTGTCTTATGGTAGCATTTGGGCTAACGCCAATTTATCAAGGTCAAAATAATA
 2350 2360 2370 2380 2390 2400

PHE ASN LYS ILE SER SER ASP VAL ILE GLY GLY ASN ALA ILE ALA THR ALA SER ALA GLU
 ATTTAATAAGATAAGTCTCTGATGTGATTGGTGGTAATGCCAATCGCTACAGCTAGCGCAGA
 2410 2420 2430 2440 2450 2460

FIG.1D.(CONTINUED)

LEU ILE VAL PRO THR PRO PHE VAL SER ASP LYS SER GIN ASN THR VAL ARG THR SER LEU
 GTTAAATTGTGCCCAACTCCATTGTGTAGTGATCAAGAGTCAAAATACAGTCCGGAACCTCCCTT
 2470 2480 2490 2500 2510 2520

PHE VAL ASP ALA ALA SER VAL TRP ASN THR LYS TRP LYS SER ASP LYS ASN GLY LEU GLU
 ATTGTGTGATGCGGCAAGTGTGTTTGGAAATACTAATAATGGGAAATCAGATAAAATGGATTAGA
 2530 2540 2550 2560 2570 2580

SER ASN VAL LEU LYS ASP LEU PRO ASP TYR GLY LYS SER SER ARG THR ARG ALA SER THR
 GAGCAATGTCCTTGAAAGACTTACCCGATTATGGCAAATCAAGCCGTA CTGCGCCCTCTAC
 2590 2600 2610 2620 2630 2640

GLY VAL GLY PHE GIN TRP GIN SER PRO SER GLY PRO VAL PHE SER TYR ALA LYS PRO
 AGGTGTCGGATTCCCAATGGCAATCTCCTAGTGGACCAAGTGGTATTTCTTATGCTAAACC
 2650 2660 2670 2680 2690 2700

ILE LYS LYS TYR GLU ASN ASP VAL GLU GIN PHE GIN PHE SER ILE GLY SER PHE
 AATTAAATAATGAAATGAAATGATGATGTCGGAACAGTTCCCAATTAGTATTGGGGGTTCTTT
 2710 2720 2730 2740 2750 2760

*** ***

CTAATAAATTGAACCTTTTTCGTCAATCAGAACTCAAAACAAACGTTCTCTGCCCTAAATTA
 2770 2780 2790 2800 2810 2820

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FIG.1D.(CONTINUED)

ATTGGGCAGAGAAATATTAAACCATCATTTAATTAAAGGATATTTATCAAAATGAAAAAC
2830 2840 2850 2860 2870 2880

ATCGCCAAAGTAACCGCACCTTGCTTTAGGTAATTGCACTTGCTTCAGGCTATGCTGCAGCT
2890 2900 2910 2920 2930 2940

GAGAGAAAAATTGCTTTTATTATAATGCAGGTTATA
2950 2960 2970

FIG.1E.

JB-1042-9-4 DNA, PAK D15
IS THE SEQUENCE BEING TRANSLATED

AAAAGGCA TTGAA A A A CAGGACA A C T T T C C C T T T T A A C C T T G A A A T A T T A G G G A A T T
10 20 30 40 50 60

A C T T A C T G G C G A T T T G T C A T T A A A T A A T T T A A G T G G G C C A A T T T C T A T T G C A A A A G G T G C
70 80 90 100 110 120

T G G T G C A T C A G C A A A T A T T G G A T T G G T G T A T T T T T A A G T T T T A T G G C A T T G A T T A G T G T
130 140 150 160 170 180 35/82

A A A T T A G G G A T T A T G A A T T T A T T T C C A T T A C C A G T A T T A G A T G G C G G T C A T T T A G T T T T
190 200 210 220 230 240

T T T A A C A A T G G A A G C T G T T A A A G G A A A A C C T G T T T C T G A G C G G G T G C A A A G C A T C T G T T A
250 260 270 280 290 300

T C G A A T T G G C G C A G C A C T G T T A T T A A G C T T A A C G G T G T T T G C A T T A T T T A A T G A T T T T T
310 320 330 340 350 360

FIG.1E.(CONTINUED)

MET LYS LYS LEU LEU ILE ALA SER LEU LEU P
 ACGTCTATAATTATATAGGATACAAATCGATGAAAAAAGTTCTTAATCGCAAGTTTATTAT
 370 380 390 400 410 420
 HE GLY THR THR THR VAL PHE ALA ALA PRO PHE VAL ALA LYS ASP ILE ARG VAL ASP G
 TCGGTACGACAAACGACTGTGTGTTTGCCCGCACCTTTGTGGCAAAAGATATTCTGCTGTGGATG
 430 440 450 460 470 480
 LY VAL GLN GLY ASP LEU GLU GLN GIN ILE ARG ALA SER LEU PRO VAL ARG ALA GLY GLN A
 GTGTTCAAGGTGACTTAGAACACAAATCCGAGCAAGTTTACCCTGTTCTGCTGGTCAGC
 490 500 510 520 530 540
 RG VAL THR ASP ASN ASP VAL ALA ASN ILE VAL ARG SER LEU PHE VAL SER GLY ARG PHE A
 GTGTAAGTACAAATGATGTGGCTAATAATTGTCCTCTCTTTATTCGTAAGTGGTCGATTCTG
 550 560 570 580 590 600
 SP ASP VAL LYS ALA HIS GIN GLU GLY ASP VAL LEU VAL SER VAL VAL ALA LYS SER I
 ATGATGTGAAAGCGCATCAAGAGGCGATGTGCTTGTGTTAGCGTTGTGGCTAAATCGA
 610 620 630 640 650 660
 LE ILE SER ASP VAL LYS ILE LYS GLY ASN SER VAL ILE PRO THR GLU ALA LEU LYS GLN A
 TCATTTCAGATGTTAAATCAAAAGGTAACTCTGTATTATTCCTCACTGAAGCACTTAAACAAA
 670 680 690 700 710 720

FIG.1E.(CONTINUED)

SN LEU ASP ALA ASN GLY PHE LYS VAL GLY ASP VAL LEU ILE ARG GLU LYS LEU ASN GLU P
 ACTTAGATGCTAACGGGTTTAAAGTTGGCGGATGTTTAAATTCGAGAAATAATTAATGAAT 780
 730 740 750 760 770

HE ALA LYS SER VAL LYS GLU HIS TYR ALA SER VAL GLY ARG TYR ASN ALA THR VAL GLU P
 TTGCCAAAAGTGTAAGAGCAGCACTATGCAAGTGTAAGTCCGCTATAACGCAACCCGTTGAAC 840
 790 800 810 820 830

RO ILE VAL ASN THR LEU PRO ASN ASN ARG ALA GLU ILE LEU ILE GIN ILE ASN GLU ASP A
 CTATTGTCAATACGCTGCGCTGCCAAATAATCGTGCTGAATAATTTAATTCAAATCAATGAAGATG 900
 850 860 870 880 890

SP LYS ALA LYS LEU ALA SER LEU THR PHE LYS GLY ASN GLU SER VAL SER SER THR L
 ATAAGCAAAATTGGCAATCACTTAACCTTCAAGGGGGAACGAATCTGTAGTAGCAGTACAT 960
 910 920 930 940 950

EU GIN GLU GIN MET GLU LEU GIN PRO ASP SER TRP TRP LYS LEU TRP GLY ASN LYS PHE G
 TACAAGAACAAATGGAAATTACAACCTGATTCCTTGGTGGAATAATTATGGGGGAAATAAATTG 1020
 970 980 990 1000 1010

LU GLY ALA GIN PHE GLU LYS ASP LEU GIN ALA ILE ARG ASP TYR TYR LEU ASN ASN GLY T
 AAGGTGCGCAATTTCGAGAAAGATCTGCAGGCAATTTCGTGATTATTTAATAATGGCT 1080
 1030 1040 1050 1060 1070

YR ALA LYS ALA GIN ILE THR LYS THR ASP VAL GIN LEU ASN ASP GLU LYS THR LYS VAL A
 ATGCCAAAGCACAAATCACTAAACGCGATGTTTCAGCTAAATGATGAAAAACAAAGTTA 1140
 1090 1100 1110 1120 1130

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FIG.1E.(CONTINUED)

SN VAL THR ILE ASP VAL ASN GLU GLY LEU GIN TYR ASP LEU ARG SER ALA ARG ILE ILE G
 ATGTAACCATGATGTAATAATGAAGGTTTACAGTATGACCTTCGTAGTGCAACGCAATTATAG
 1150 1160 1170 1180 1190 1200

LY ASN LEU GLY GLY MET SER ALA GLU LEU LEU GLU PRO LEU LEU SER ALA LEU HIS LEU ASN A
 GTAATCTGGGAGGTATGTCCTGCGGAGCCTTGAAACCTTTACTTTTCAGCATTAATTAAATG
 1210 1220 1230 1240 1250 1260

SP THR PHE ARG ARG SER ASP ILE ALA ASP VAL GLU ASN ALA ILE LYS ALA LYS LEU GLY G
 A T A C T T C C G C C G T A G T G A T A T T G C A G A T G T A G A A A T T A A G C A A A C T T G G G G
 1270 1280 1290 1300 1310 1320

LU ARG GLY TYR GLY ASN THR THR VAL ASN SER VAL PRO ASP PHE ASP ALA ASN LYS T
 A C G A G G T T A C G G T A A C A C A C A C A G T A A A T T C T G T A C C T G A T T T G A C G A T G C A A T A A A
 1330 1340 1350 1360 1370 1380

HR LEU ALA ILE THR PHE VAL VAL ASP ALA GLY ARG ARG LEU THR VAL ARG GIN LEU ARG P
 C A T T A G C C G A T A A C C T T T G T T G T T G A T G C T G G A C G A C G T T T A A C T G T T C G C C A A C T T C G C T
 1390 1400 1410 1420 1430 1440

HE GLU GLY ASN THR VAL SER ALA ASP SER THR LEU ARG GIN GLU MET ARG GIN GIN GLU G
 T T G A A G G A A A T A C C G T T T C T G C T G A T A G T A C T T T A C G T C A G G A A A T G C G A C A C A A G A A G
 1450 1460 1470 1480 1490 1500

FIG.1E.(CONTINUED)

LY THR TRP TRP ASN SER GIN LEU VAL GLU LEU GLY LYS ILE ARG LEU ASP ARG THR GLY P
 GAACTTGGGTATAATTCAAAATTAGTTGAGTTAGGAAATAATTCGCTTAGATCGGTACAGGTT
 1510 1520 1530 1540 1550 1560

HE PHE GLU THR VAL GLU ASN ARG ILE ASP PRO ILE ASN GLY SER ASN ASP GLU VAL ASP V
 TCTTCGAAACAGTTGAAACCGAATTGATCCTATCAATGGTAGCAATGATGAAGTGGATG
 1570 1580 1590 1600 1610 1620

AL VAL TYR LYS VAL LYS GLU ARG ASN THR GLY SER ILE ASN PHE GLY ILE TYR GLY T
 TCGTATATAAAGTCAAGAAACGTAACACGGGTTAGTATCAAACTTTGGTATTGGTTACGGTA
 1630 1640 1650 1660 1670 1680

HR GLU SER GLY ILE SER TYR GIN THR SER ILE LYS GIN ASP ASN PHE LEU GLY THR GLY A
 CAGAGAGTGGTATCAGTTATCAACAACAAGTATTAAACAAGATAATTCTTTGGGAACAGGGG
 1690 1700 1710 1720 1730 1740

LA ALA VAL SER ILE ALA GLY THR LYS ASN ASP TYR GLY THR SER VAL ASN LEU GLY TYR T
 CGGCAGTAGTATAGCTGGTACGAAATAATGATTATGGTACGAGTGTCATAATTGGGTATA
 1750 1760 1770 1780 1790 1800

HR GLU PRO TYR PHE THR LYS ASP GLY VAL SER LEU GLY GLY ASN ILE PHE PHE GLU ASN T
 CCGAACCCCTATTCTACTAAAGATGGTGTAAAGTCTTGGTGGAAATAATTTCTTTGAAACT
 1810 1820 1830 1840 1850 1860

FIG.1E.(CONTINUED)

YR ASP ASN SER LYS SER ASP THR SER SER ASN TYR LYS ARG THR THR TYR GLY SER ASN V
 ACGATAACTCTAAAGTGATACATCCTCTAATAAGCGTACGACTTATGGAAAGTAATG 1870 1880 1890 1900 1910 1920
 AL THR LEU GLY PHE PRO VAL ASN GLU ASN ASN SER TYR TYR VAL GLY LEU GLY HIS THR T
 TTACTTTAGGTTTCCCTGTAAATGAAATAAATCCTATTAATGTTAGGATTAAGCCATACCT 1930 1940 1950 1960 1970 1980
 YR ASN LYS ILE SER ASN PHE ALA LEU GLU TYR ASN ARG ASN LEU TYR ILE GLN SER MET L
 ATAATAAATTAGTAACCTTTGCTCTAGATAATAACCGTAATTATATATCAATCAATGA 1990 2000 2010 2020 2030 2040
 YS PHE LYS GLY ASN GLY ILE LYS THR ASN ASP PHE ASP PHE SER PHE GLY TRP ASN TYR A
 AATTAAAGGTAATGGCATTAACAACAATGACTTTTGATTTCTTTGGTTGGAACTATA 2050 2060 2070 2080 2090 2100
 SN SER LEU ASN ARG GLY TYR PHE PRO THR LYS GLY VAL LYS ALA SER LEU GLY ARG V
 ACAGCCTTAATAGAGGCTATTTCCTCACTAAAGGGTTAAAGCAAGTCTTGGTGGACGAG 2110 2120 2130 2140 2150 2160
 AL THR ILE PRO GLY SER ASP ASN LYS TYR TYR LYS LEU SER ALA ASP VAL GLN GLY PHE T
 TACTATTCCAGGTTCTGTGATAACAATACTACTACAACCTAAGTGCAAGTACAGGGTTTCT 2170 2180 2190 2200 2210 2220
 YR PRO LEU ASP ARG ASP HIS ARG TRP VAL VAL SER ALA LYS ALA SER ALA GLY TYR ALA A
 ACCCATTAGACAGAGATCACCGCTGGGTTGTATCTGCAAAAGCACTCGCAGGATATGCAA 2230 2240 2250 2260 2270 2280

40/02

FIG.1E.(CONTINUED)

SN GLY PHE GLY ASN LYS ARG LEU PRO PHE TYR GLN THR TYR THR ALA GLY GLY ILE GLY S
 ATGGTTT TGGAAACAAGCGTTTACCGTTCTATCAAACTTATACAGCGGTGGCATTTGGTT 2300 2330 2340

ER LEU ARG GLY PHE ALA TYR GLY SER ILE GLY PRO ASN ALA ILE TYR ALA GLU HIS GLY A
 CATTACCGCGGTTT TGGCTTATGGTAGTATTGGGCCCTAATGCAATTATTATGCCCGAACAATGGTA 2350 2370 2380 2390 2400

SN GLY THR PHE ASN LYS ILE SER SER ASP VAL ILE GLY GLY ASN ALA ILE THR THR ALA S
 ATGGTACTTTTAAATAAGATAAGTCTGATGTGATTGGTGTTAATGCCAATCACAAC TGGCGA 2410 2420 2430 2440 2450 2460 41/82

ER ALA GLU LEU ILE VAL PRO THR PRO PHE VAL SER ASP LYS SER GLN ASN THR VAL ARG T
 GTGCAGAACTTATTGTACCAACTCCATTCTGATGTGAGTGATAAAGCCCAAATAACAGTCCGAA 2470 2480 2490 2500 2510 2520

HR SER LEU PHE VAL ASP ALA ALA SER VAL TRP ASN THR LYS TRP LYS SER ASP LYS ASN G
 CCTCCCTATTGTGTTGATGCGGCAAGTGTTTGGGAATACTAAATGGAAATCAGATAAATAATG 2530 2540 2550 2560 2570 2580

LY LEU GLU SER LYS VAL LEU LYS ASP LEU PRO ASP TYR GLY LYS SER SER ARG ILE ARG A
 GATTAGAGAGCAAGGTC TTGAAAGACTTACCTGATTATGGCAATAATCAAGCCGTTATTCGCG 2590 2600 2610 2620 2630 2640

FIG.1E.(CONTINUED)

LA SER THR GLY VAL GLY PHE GLN TRP GLN SER PRO ILE GLY PRO LEU VAL PHE SER TYR A
 CCTCTACAGGTGTCGGAATTCCAATGGCAATCTCCTATTGGACCAATTGGGTAATTTCTTATG
 2650 2660 2670 2680 2690 2700

LA LYS PRO ILE LYS LYS TYR GLU ASN ASP VAL GLU GLN PHE SER ILE GLY G
 CTAACCAATTAAATAATATGAATAATGATGATGTCGAACAGTTCCAAATTAGTATTGGGG
 2710 2720 2730 2740 2750 2760

LY SER PHE *** ***

GCTCTTCTAATAAATTGAACCTTTTTCGTCAACAGAAACGACGTTCTCTGCC
 2770 2780 2790 2800 2810 2820

TAAATTGAATTGGGCAGAGAAATAATTAAACCCATCATTTAATAAGGATATTTATCAAA
 2830 2840 2850 2860 2870 2880

GAAACAATCGCAAAAGTAACCGCACCTTGCTTTAGGTTTTCACCTTGCTTCAGGCTATGC
 2890 2900 2910 2920 2930 2940

TTCCGCTGAAGAAATAATTGCTTTCAATTAAATGCAGGTTATATTTTCAA
 2950 2960 2970 2980

42/82

FIG.1F.

1. cad15 (1-2949)
3. minnad15 (1-2953)
2. eagand15 (1-2984)
4. pakd15 (1-2989)
5. sb33d15 (1-2974)

cad15 1

minnad15 1

eagand15 1

pakd15 1

sb33d15 1

consensus

cad15 1

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ACAGGACAGCTTTCCCTTTTAAACCTTGAAAAATATTAGGAAATTA
|||||
1 aaaggcattgaaaaaacaggacagctttcccttttaacaccttgaaaataattagggaatta
|||||
1 GGCATTGAAAAAACAGGACAGCTTTCCCTTTTAAACCTTGAAAAATATTAGGAAATTA
|||||
1 aaaggcattgaaaaaacaggacagctttcccttttaacaccttgaaaataattagggaatta

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FIG. 1F.(CONTINUED)

cad15	1	gATTAC
minnad15	128	TTTAGGGATTATGAATTATTTCCTATTACCAGTATTAGATGGCGGTCAATTAGTTTTTTTA
eagand15	168	TTTAGGGATTATGAATTATTTCCTATTACCAGTATTAGATGGCGGTCAATTAGTTTTTTTA
pakd15	184	TTTAGGGATTATGAATTATTTCCTATTACCAGTATTAGATGGCGGTCAATTAGTTTTTTTA
sb33d15	180	TTTAGGGATTATGAATTATTTCCTATTACCAGTATTAGATGGCGGTCAATTAGTTTTTTTA
consensus		tttagggattatgaattatttccATTACcagtagatttagatggcgggtcatbtagttttttta

cad15	7	
minnad15	189	ACAAATGGAAGCTGTTAAAGGAAAACCTGTTTCTGAGCGGGTGCAAAGCATCTGTTATCGAA
eagand15	229	ACAAATGGAAGCTGTTAAAGGAAAACCTGTTTCTGAGCGGGTGCAAAGCATCTGTTATCGAA
pakd15	245	ACAAATGGAAGCTGTTAAAGGAAAACCTGTTTCTGAGCGGGTGCAAAGCATCTGTTATCGAA
sb33d15	241	ACAAATGGAAGCTGTTAAAGGAAAACCTGTTTCTGAGCGGGTGCAAAGCATCTGTTATCGAA

FIG.1F.(CONTINUED)

consensus acaatggaagctgttaaaggaaaacctgttctgagcgggtgcaaaagcatctgttatcgaa

cad15 7 gccAAGCTTAACGGTGTGTCATTATTTAAATGATTTTACGTCT
 minnad15 250 TTGGCGCAGCACTGTTATTAAGCTTAACGGTGTGTCATTATTTAAATGATTTTACGTCT
 eagand15 290 TTGGCGCAGCACTGTTATTAAGCTTAACGGTGTGTCATTATTTAAATGATTTTACGTCT
 pakd15 306 TTGGCGCAGCACTGTTATTAAGCTTAACGGTGTGTCATTATTTAAATGATTTTACGTCT
 sb33d15 302 TTGGCGCAGCACTGTTATTAAGCTTAACGGTGTGTCATTATTTAAATGATTTTACGTCT
 consensus ttggcgcagcactgttattAAGCTTAACGGTGTGTCATTATTTAAATGATTTTACGTCT

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cad15 52 ATAAATTATATAGGATACAATCGATGAATAAACTTCTAATCGCAAGTTATATATTCGGTAC
 minnad15 311 ATAAATTATATAGGATACAATCGATGAATAAACTTCTAATCGCAAGTTATATATTCGGTAC
 eagand15 351 ATAAATTATATAGGATACAATCGATGAATAAACTTCTAATCGCAAGTTATATATTCGGTAC
 pakd15 367 ATAAATTATATAGGATACAATCGATGAATAAACTTCTAATCGCAAGTTATATATTCGGTAC

FIG.1F.(CONTINUED)

sb333d15 363 ATAATTATATAGGATACAATCGATGAAAAAAGTTCTAATCGCAAGTTTATTATTCGGTAc
consensus ATAATTATATAGGATACAATCGATGAAAAAAGTTCTAATCGCAAGTTTATTATTCGGTAc

cad15 113 GACAACGACTGTGTTGCCGCACCTTTTGTGGCAAAGATATTCGTGTGGATGGTGTCAA
|||||
minnad15 372 GACAACGACTGTGTTGCCGCACCTTTTGTGGCAAAGATATTCGTGTGGATGGTGTCAA
|||||
eagand15 412 GACAACGACTGTGTTGCCGCACCTTTTGTGGCAAAGATATTCGTGTGGATGGTGTCAA
|||||
pakd15 428 GACAACGACTGTGTTGCCGCACCTTTTGTGcCAAAGATATTCGTGTGGATGGTGTCAA
|||||
sb333d15 424 GACAACGACTGTGTTGCCGCACCTTTTGTGgCAAAGATATTCGTGTGGATGGTGTCAA
|||||

47/82

consensus GACAACGACTGTGTTGCCGCACCTTTTGTGgCAAAGATATTCGTGTGGATGGTGTCAA

cad15 174 GGTGACTTAGAACAAACAATCCGAGCAAGTTTACCTGTTCGTGCCGGTCAGCGTGTGACTG
|||||
minnad15 433 GGTGACTTAGAACAAACAATCCGAGCAAGTTTACCTGTTCGTGCCGGTCAGCGTGTGACTG
|||||
eagand15 473 GGTGACTTAGAACAAACAATCCGAGCAAGTTTACCTGTTCGTGCCGGTCAGCGTGTGACTG
|||||

FIG.1F.(CONTINUED)

pakd15 489 GGTGACTTAGAACAAACAAATCCGAGCAAGTTTACCTGTTCGTGCTGGTCAGCGTGTGACTG
 |||||
 sb33d15 485 GGTGACTTAGAACAAACAAATCCGAGCAAGTTTACCTGTTCGTGCTGGTCAGCGTGTGACTG
 |||||
 consensus GGTGACTTAGAACAAACAAATCCGAGCAAGTTTACCTGTTCGTGCTGGTCAGCGTGTGACTG

cad15 235 ACAATGATGTGGCTAATAATTGTCCGCTCTTTATTTCGTAAGTGGTCGATTCGATGTGAA
 |||||
 minnad15 494 ACAATGATGTGGCTAATAATTGTCCGCTCTTTATTTCGTAAGTGGTCGATTCGATGTGAA
 |||||
 eagand15 534 ACAATGATGTGGCTAATAATTGTCCGCTCTTTATTTCGTAAGTGGTCGATTCGATGTGAA
 |||||
 pakd15 550 ACAATGATGTGGCTAATAATTGTCCGCTCTTTATTTCGTAAGTGGTCGATTCGATGTGAA
 |||||
 sb33d15 546 ACAATGATGTGGCTAATAATTGTCCGCTCTTTATTTCGTAAGTGGTCGATTCGATGTGAA
 |||||

40/82

consensus ACAATGATGTGGCTAATAATTGTCCGCTCTTTATTTCGTAAGTGGTCGATTCGATGTGAA

cad15 296 AGCGCATCAAGAAAGCGGATGTGCTTGTGTTAGCGTTGTGGCTAAATCGATCATTTTCAGAT
 |||||
 minnad15 555 AGCGCATCAAGAAAGCGGATGTGCTTGTGTTAGCGTTGTGGCTAAATCGATCATTTTCAGAT
 |||||

FIG.1F.(CONTINUED)

eagand15	595	AGCGCATCAAGAAGCGCATGTGCTTGTGTAGCGTTGTGGCTAAATCGATCATTTTCAGAT	
pakd15	611	AGCGCATCAAGAAGCGCATGTGCTTGTGTAGCGTTGTGGCTAAATCGATCATTTTCAGAT	
sb33d15	607	AGCGCATCAAGAAGCGCATGTGCTTGTGTAGCGTTGTGGCTAAATCGATCATTTTCAGAT	
consensus		AGCGCATCAAGAAGCGCATGTGCTTGTGTAGCGTTGTGGCTAAATCGATCATTTTCAGAT	
cad15	357	GTAAATAATCAAGGTAACCTCTGTTATTCCTCACTGAAGCACTTAAACAAACTTAGATGCTA	49/82
minnad15	616	GTAAATAATCAAGGTAACCTCTGTTATTCCTCACTGAAGCACTTAAACAAACTTAGATGCTA	
eagand15	656	GTAAATAATCAAGGTAACCTCTGTTATTCCTCACTGAAGCACTTAAACAAACTTAGATGCTA	
pakd15	672	GTAAATAATCAAGGTAACCTCTGTTATTCCTCACTGAAGCACTTAAACAAACTTAGATGCTA	
sb33d15	668	GTAAATAATCAAGGTAACCTCTATTATTCCTCACTGAAGCACTTAAACAAACTTAGATGCTA	
consensus		GTAAATAATCAAGGTAACCTCTATTATTCCTCACTGAAGCACTTAAACAAACTTAGATGCTA	
cad15	418	ACGGGTTTAAAGTTGGCGATGTTTTAATTCGAGAAAAATTAAATGAATTTGCCAAAAAGTGT	

FIG. 1F.(CONTINUED)

minnad15	677	ACGGGTTTAAAGTTGGCGATGTTTAAATTCGAGAAAAATTAAATGAATTTGCCAAAAAGTGT	
eagand15	717	ACGGGTTTAAAGTTGGCGATGTTTAAATTCGAGAAAAATTAAATGAATTTGCCAAAAAGTGT	
pakd15	733	ACGGGTTTAAAGTTGGCGATGTTTAAATTCGAGAAAAATTAAATGAATTTGCCAAAAAGTGT	
sb33d15	729	ACGGGTTTAAAGTTGGCGATGTTTAAATTCGAGAAAAATTAAATGAATTTGCCAAAAAGTGT	
consensus		ACGGGTTTAAAGTTGGCGATGTTTAAATTCGAGAAAAATTAAATGAATTTGCCAAAAAGTGT	
			50/82
cad15	479	AAAAGAGCACTATGCAAGTGTAAGTCGCTATAACGCAACAGTTGAACCTATTGTCAATACG	
minnad15	738	AAAAGAGCACTATGCAAGTGTAAGTCGCTATAACGCAACAGTTGAACCTATTGTCAATACG	
eagand15	778	AAAAGAGCACTATGCAAGTGTAAGTCGCTATAACGCAACAGTTGAACCTATTGTCAATACG	
pakd15	794	AAAAGAGCACTATGCAAGTGTAAGTCGCTATAACGCAACAGTTGAACCTATTGTCAATACG	
sb33d15	790	AAAAGAGCACTATGCAAGTGTAAGTCGCTATAACGCAACAGTTGAACCTATTGTCAATACG	
consensus		AAAAGAGCACTATGCAAGTGTAAGTCGCTATAACGCAACAGTTGAACCTATTGTCAATACG	

FIG.1F.(CONTINUED)

cad15 540 CTACCAAAATAATCGCGCTGAAATTTTAATTCAAATCAATGAAGATGATAAAGCAAAATTGG
 minnad15 799 CTACCAAAATAATCGCGCTGAAATTTTAATTCAAATCAATGAAGATGATAAAGCAAAATTGG
 eagand15 839 CTACCAAAATAATCGCGCTGAAATTTTAATTCAAATCAATGAAGATGATAAAGCAAAATTGG
 pakd15 855 CTGCCAAAATAATCGCGCTGAAATTTTAATTCAAATCAATGAAGATGATAAAGCAAAATTGG
 sb33d15 851 CTaCCAAAATAATCGCGCTGAAATTTTAATTCAAATCAATGAAGATGATAAAGCCAAATTGG

51/82

consensus CTaCCAAAATAATCGCGCTGAAATTTTAATTCAAATCAATGAAGATGATAAAGCaAAATTGG
 cad15 601 CATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGTACATTACAAGAACAAATGGA
 minnad15 860 CATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGTACATTACAAGAACAAATGGA
 eagand15 900 CATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGTACATTACAAGAACAAATGGA
 pakd15 916 CATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGTACATTACAAGAACAAATGGA
 sb33d15 912 CATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGTACATTACAAGAACAAATGGA
 consensus CATCATTAACCTTCAAGGGGAACGAATCTGTAGTAGCAGTACATTACAAGAACAAATGGA

FIG.1F.(CONTINUED)

cad15 662 ATTACAACCTGATTCTTGGTGGAATATATGGGAAATAAAATTTGAAGGTGCGCAATTCGAG
 |||||
 minnad15 921 ATTACAACCTGATTCTTGGTGGAATATATGGGAAATAAAATTTGAAGGTGCGCAATTCGAG
 |||||
 eagand15 961 ATTACAACCTGATTCTTGGTGGAATATATGGGAAATAAAATTTGAAGGTGCGCAATTCGAG
 |||||
 pakd15 977 ATTACAACCTGATTCTTGGTGGAATATATGGGAAATAAAATTTGAAGGTGCGCAATTCGAG
 |||||
 sb33d15 973 ATTACAACCTGATTCTTGGTGGAATATATGGGAAATAAAATTTGAAGGTGCGCAATTCGAG
 |||||

52/82

consensus ATTACAACCTGATTCTTGGTGGAATATATGGGAAATAAAATTTGAAGGTGCGCAATTCGAG
 |||||
 cad15 723 AAAGATTTCAGTCAATTTCGTGATTATTTAAATAATGGCTATGCCAAAGCACAAAATTA
 |||||
 minnad15 982 AAAGATTTCAGTCAATTTCGTGATTATTTAAATAATGGCTATGCCAAAGCACAAAATTA
 |||||
 eagand15 1022 AAAGATTTCAGTCAATTTCGTGATTATTTAAATAATGGCTATGCCAAAGCACAAAATTA
 |||||
 pakd15 1038 AAAGATcTGCAGGCAATTTCGTGATTATTTAAATAATGGCTATGCCAAAGCACAAAATCA
 |||||
 sb33d15 1034 AAAGATtTGCAGGCAATTTCGTGATTATTTAAATAATGGCTATGCCAAAGCACAAAATCA
 |||||
 consensus AAAGATtTGCAGtCAATTTCGTGATTATTTAAATAATGGCTATGCCAAAGCACAAAATtA

FIG.1F.(CONTINUED)

cad15 784 CTAAACGGATGTTTCAGCTAAATGATGAAAAACAAAAGTTAATGTAACCATTTGATGTAAA
 |||||
 minnad15 1043 CTAAACGGATGTTTCAGCTAAATGATGAAAAACAAAAGTTAATGTAACCATTTGATGTAAA
 |||||
 eagand15 1083 CTAAACGGATGTTTCAGCTAAATGATGAAAAACAAAAGTTAATGTAACCATTTGATGTAAA
 |||||
 pakd15 1099 CTAAACGGATGTTTCAGCTAAATGATGAAAAACAAAAGTTAATGTAACCATTTGATGTAAA
 |||||
 sb33d15 1095 CTAAAGCGGATGTTTCAGCTAAATGATGAAAAACAAAAGTTAATGTAACCATTTGATGTAAA
 |||||

53/82

consensus CTAAACGGATGTTTCAGCTAAATGATGAAAAACAAAAGTTAATGTAACCATTTGATGTAAA
 |||||
 cad15 845 TGAAGGTTTACAGTATGACCTTCGTTAGTGCACGCATTATAGGTAATCTGGGAGGTATGTCT
 |||||
 minnad15 1104 TGAAGGTTTACAGTATGACCTTCGTTAGTGCACGCATTATAGGTAATCTGGGAGGTATGTCT
 |||||
 eagand15 1144 TGAAGGTTTACAGTATGACCTTCGTTAGTGCACGCATTATAGGTAATCTGGGAGGTATGTCT
 |||||
 pakd15 1160 TGAAGGTTTACAGTATGACCTTCGTTAGTGCACGCATTATAGGTAATCTGGGAGGTATGTCT
 |||||
 sb33d15 1156 TGAAGGTTTACAGTATGACCTTCGTTAGTGCACGCATTATAGGTAATCTGGGAGGTATGTCT
 |||||
 consensus TGAAGGTTTACAGTATGACCTTCGTTAGTGCACGCATTATAGGTAATCTGGGAGGTATGTCT

FIG.1F.(CONTINUED)

cad15 906 GCCGAGCTTGAACCTTACTTTCAGCATTACATTTAAATGATACTTCCGCCGCTAGTGATA
 |||||
 minnad15 1165 GCCGAGCTTGAACCTTACTTTCAGCATTACATTTAAATGATACTTCCGCCGCTAGTGATA
 |||||
 eagand15 1205 GCCGAGCTTGAACCTTACTTTCAGCATTACATTTAAATGATACTTCCGCCGCTAGTGATA
 |||||
 pakd15 1221 GCCGAGCTTGAACCTTACTTTCAGCATTACATTTAAATGATACTTCCGCCGCTAGTGATA
 |||||
 sb33d15 1217 GCCGAGCTTGAACCTTACTTTCAGCATTACATTTAAATGATACTTCCGCCGCTAGTGATA
 |||||
 consensus GCCGAGCTTGAACCTTACTTTCAGCATTACATTTAAATGATACTTCCGCCGCTAGTGATA

5 4/82

cad15 967 TTGCAGATGTAGAAAATGCAATTAAAGCAAAACTTGGAGAACGCGGTTACGGTAGCGCAAC
 |||||
 minnad15 1226 TTGCAGATGTAGAAAATGCAATTAAAGCAAAACTTGGAGAACGCGGTTACGGTAGCGCAAC
 |||||
 eagand15 1266 TTGCAGATGTAGAAAATGCAATTAAAGCAAAACTTGGAGAACGCGGTTACGGTAGCGCAAC
 |||||
 pakd15 1282 TTGCAGATGTAGAAAATGCAATTAAAGCAAAACTTGGGGAACGAGGTTACGGTAACACAAC
 |||||
 sb33d15 1278 TTGCAGATGTAGAAAATGCAATTAAAGCAAAACTTGGGGAACGAGGTTACGGTAACACAAC
 |||||
 consensus TTGCAGATGTAGAAAATGCAATTAAAGCAAAACTTGGGAACGCGGTTACGGTAGCGCAAC

FIG.1F.(CONTINUED)

cad15	1028	GGTAAATTCA	GACCTGATTT	GATGATGCA	ATAAAACAT	TAGCGATA	AACCCCTG	TTGTTGTT
minnad15	1287	GGTAAATTCA	GACCTGATTT	GATGATGCA	ATAAAACAT	TAGCGATA	AACCCCTG	TTGTTGTT
eagand15	1327	GGTAAATTCA	GACCTGATTT	GATGATGCA	ATAAAACAT	TAGCGATA	AACCCCTG	TTGTTGTT
pakd15	1343	AGTAAATTCT	GTACCTGATTT	TGACGATG	CAATAAAAC	ATTAGCGATA	AACCTTTG	TTGTTGTT
sb33d15	1339	AGTAAATTCT	GTACCTGATTT	TGACGATG	CAATAAAAC	ATTAGCGATA	AACCTTTG	TTGTTGTT
consensus		GGTAAATTCA	GACCTGATTT	TGATGATG	CAATAAAAC	ATTAGCGATA	AACCCCTG	TTGTTGTT
		5/82						
cad15	1089	GATGCTGG	ACGACGTT	TAACTGTT	CGCCAACT	TCGCTTTG	AAGGAAAT	ACCGTTTCTGCTG
minnad15	1348	GATGCTGG	ACGACGTT	TAACTGTT	CGCCAACT	TCGCTTTG	AAGGAAAT	ACCGTTTCTGCTG
eagand15	1388	GATGCTGG	ACGACGTT	TAACTGTT	CGCCAACT	TCGCTTTG	AAGGAAAT	ACCGTTTCTGCTG
pakd15	1404	GATGCTGG	ACGACGTT	TAACTGTT	CGCCAACT	TCGCTTTG	AAGGAAAT	ACCGTTTCTGCTG
sb33d15	1400	GATGCTGG	ACGACGTT	TAACTGTT	CaCCAAC	TTTCGCTTTG	AAGGAAAT	ACCGTTTCTGCTG
consensus		GATGCTGG	ACGACGTT	TAACTGTT	CGCCAACT	TCGCTTTG	AAGGAAAT	ACCGTTTCTGCTG

FIG.1F.(CONTINUED)

cad15	1150	ATAGCACTTTACGTCAGGAAATGCGCCAAACAAGAAGGAAC TTGGTATAATTCACAATTAGT	
minnad15	1409	ATAGCACTTTACGTCAGGAAATGCGCCAAACAAGAAGGAAC TTGGTATAATTCACAATTAGT	
eagand15	1449	ATAGCACTTTACGTCAGGAAATGCGCCAAACAAGAAGGAAC TTGGTATAATTCACAATTAGT	
pakd15	1465	ATAGTACTTTACGTCAGGAAATGCGCAACAAGAAGGAAC TTGGTATAATTCACAATTAGT	
sb33d15	1461	ATAGTACTTTACGTCAGGAAATGCGCCAAACAAGAAGGAAC TTGGTATAATTCACAATTAGT	
consensus		ATAGCACTTTACGTCAGGAAATGCGCCAAACAAGAAGGAAC TTGGTATAATTCACAATTAGT	5
			6/82
cad15	1211	TGAGTTAGGAAAAAATTCGCTTAGATCGTACAGGTTTCTTCGAAACAGTCGAAAACCGGAATT	
minnad15	1470	TGAGTTAGGAAAAAATTCGCTTAGATCGTACAGGTTTCTTCGAAACAGTCGAAAACCGGAATT	
eagand15	1510	TGAGTTAGGAAAAAATTCGCTTAGATCGTACAGGTTTCTTCGAAACAGTCGAAAACCGGAATT	
pakd15	1526	TGAGTTAGGAAAAAATTCGCTTAGATCGTACAGGTTTCTTCGAAACAGTTGAAAACCGGAATT	
sb33d15	1522	TGAGTTAGGAAAAAATTCGCTTAGATCGTACAGGTTTCTTCGAAACAGTTGAAAACCGGAATT	
consensus		TGAGTTAGGAAAAAATTCGCTTAGATCGTACAGGTTTCTTCGAAACAGTcGAAAACCGGAATT	

FIG.1F.(CONTINUED)

cad15	1272	GATCCTATCAATGGTAGTAATGATGAAGTGGATGTCGTATATAAAGTCAAAGAACGTAACA	
minnad15	1531	GATCCTATCAATGGTAGTAATGATGAAGTGGATGTCGTATATAAAGTCAAAGAACGTAACA	
eagand15	1571	GATCCTATCAATGGTAGTAATGATGAAGTGGATGTCGTATATAAAGTCAAAGAACGTAACA	
pakd15	1587	GATCCTATCAATGGTAGCAATGATGAAGTGGATGTCGTATATAAAGTCAAAGAACGTAACA	
sb33d15	1583	GATCCTATCAATGGTAGCAATGATGAAGTGGATGTCGTATATAAAGTCAAAGAACGTAACA	
consensus		GATCCTATCAATGGTAGTAATGATGAAGTGGATGTCGTATATAAAGTCAAAGAACGTAACA	57/82
cad15	1333	CGGGTAGTATCAACTTTGGTATTGGTTACGGTACAGAGAGTGGTATTAGTTATCAAGCAAG	
minnad15	1592	CGGGTAGTATCAACTTTGGTATTGGTTACGGTACAGAGAGTGGTATTAGTTATCAAGCAAG	
eagand15	1632	CGGGTAGTATCAACTTTGGTATTGGTTACGGTACAGAGAGTGGTATTAGTTATCAAGCAAG	
pakd15	1648	CGGGTAGTATCAACTTTGGTATTGGTTACGGTACAGAGAGTGGTATCAGTTATCAAAaCAAG	
sb33d15	1644	CGGGTAGTATCAACTTTGGTATTGGTTACGGTACAGAGAGTGGTATCAGTTATCAAGCAAG	
consensus		CGGGTAGTATCAACTTTGGTATTGGTTACGGTACAGAGAGTGGTATCAGTTATCAAGCAAG	

FIG. 1F.(CONTINUED)

cad15	1394	TGTTAAACAAGATAATTCTTGGGAACAGGGCGGCAGTAAGTATAGCTGGTACGAAAAAT	
minnad15	1653	TGTTAAACAAGATAATTCTTGGGAACAGGGCGGCAGTAAGTATAGCTGGTACGAAAAAT	
eagand15	1693	TGTTAAACAAGATAATTCTTGGGAACAGGGCGGCAGTAAGTATAGCTGGTACGAAAAAT	
pakd15	1709	TaTTAAACAAGATAATTCTTGGGAACAGGGCGGCAGTAAGTATAGCTGGTACGAAAAAT	
sb33d15	1705	TgTcAAACAAGATAATTCTTGGGAACAGGGCGGCAGTAAGTATAGCTGGTACGAAAAAT	
consensus		TgTtAAACAAGATAATTCTTGGGAACAGGGCGGCAGTAAGTATAGCTGGTACGAAAAAT	08/82
cad15	1455	GATTATGGTACGAGTGTCAATTTGGGTTATACCGAGCCCTATTTTACTAAAGATGGTGTA	
minnad15	1714	GATTATGGTACGAGTGTCAATTTGGGTTATACCGAGCCCTATTTTACTAAAGATGGTGTA	
eagand15	1754	GATTATGGTACGAGTGTCAATTTGGGTTATACCGAGCCCTATTTTACTAAAGATGGTGTA	
pakd15	1770	GATTATGGTACGAGTGTCAATTTGGGTTATACCGAaCCCTATTTTACTAAAGATGGTGTA	
sb33d15	1766	GATTATGGTACGAGTGTCAATTTGGGTTATACCGAGCCCTATTTTACTAAAGATGGTGTA	
consensus		GATTATGGTACGAGTGTCAATTTGGGTTATACCGAGCCCTATTTTACTAAAGATGGTGTA	

FIG.1F.(CONTINUED)

cad15	1516	GTCTTGGTGGAAATGTTTCTTTGAAAACTACGATAACTCTAAAAGTGATACATCCTCTTAA	
minnad15	1775	GTCTTGGTGGAAATGTTTCTTTGAAAACTACGATAACTCTAAAAGTGATACATCCTCTTAA	
eagand15	1815	GTCTTGGTGGAAATGTTTCTTTGAAAACTACGATAACTCTAAAAGTGATACATCCTCTTAA	
pakd15	1831	GTCTTGGTGGAAATGTTTCTTTGAAAACTACGATAACTCTAAAAGTGATACATCCTCTTAA	
sb33d15	1827	GTCTTGGTGGAAATGTTTCTTTGAAAACTACGATAACTCTAAAAGTGATACATCCTCTTAA	
consensus		GTCTTGGTGGAAATGTTTCTTTGAAAACTACGATAACTCTAAAAGTGATACATCCTCTTAA	59/82
cad15	1577	CTATAAGCGTACGACTTACGGAAGTAATGTTACTTTAGGTTTCCCTGTAAAATGAAAATAAC	
minnad15	1836	CTATAAGCGTACGACTTACGGAAGTAATGTTACTTTAGGTTTCCCTGTAAAATGAAAATAAC	
eagand15	1876	CTATAAGCGTACGACTTACGGAAGTAATGTTACTTTAGGTTTCCCTGTAAAATGAAAATAAC	
pakd15	1892	CTATAAGCGTACGACTTATGGAAGTAATGTTACTTTAGGTTTCCCTGTAAAATGAAAATAAC	
sb33d15	1888	CTATAAGCGTACGACTTATGGAAGTAATGTTACTTTAGGTTTCCCTGTAAAATGAAAATAAC	
consensus		CTATAAGCGTACGACTTACGGAAGTAATGTTACTTTAGGTTTCCCTGTAAAATGAAAATAAC	

FIG.1F.(CONTINUED)

cad15	1638	TCCTATTATGTAGGATTAGGTCATACCTATAATAAAATTAGTAACTTTTGCTCTAGAAATATA
minnad15	1897	TCCTATTATGTAGGATTAGGTCATACCTATAATAAAATTAGTAACTTTTGCTCTAGAAATATA
eagand15	1937	TCCTATTATGTAGGATTAGGTCATACCTATAATAAAATTAGTAACTTTTGCTCTAGAAATATA
pakd15	1953	TCCTATTATGTAGGATTAGGCCATACCTATAATAAAATTAGTAACTTTTGCTCTAGAAATATA
sb33d15	1949	TCCTATTATGTAGGATTAGGCCATACCTATAATAAAATTAGTAACTTTTGCTCTAGAAATATA
consensus		TCCTATTATGTAGGATTAGGTCATACCTATAATAAAATTAGTAACTTTTGCTCTAGAAATATA
		60/82
cad15	1699	ACCGTAAATTTATATATTCAATCAATGAAATTTAAAGGTAATGGCATTTAAACAAATGACTT
minnad15	1958	ACCGTAAATTTATATATTCAATCAATGAAATTTAAAGGTAATGGCATTTAAACAAATGACTT
eagand15	1998	ACCGTAAATTTATATATTCAATCAATGAAATTTAAAGGTAATGGCATTTAAACAAATGACTT
pakd15	2014	ACCGTAAATTTATATATTCAATCAATGAAATTTAAAGGTAATGGCATTTAAACAAATGACTT
sb33d15	2010	ACCGTAAATTTATATATTCAATCAATGAAATTTAAAGGTAATGGCATTTAAACAAATGACTT
consensus		ACCGTAAATTTATATATTCAATCAATGAAATTTAAAGGTAATGGCATTTAAACAAATGACTT

FIG.1F.(CONTINUED)

cad15	1760	TGATTTTCTTTTGGTTGGAACTATAACAGCCCTTAATAGAGGCTATTTCCCAACATAAAGGG	
minnad15	2019	TGATTTTCTTTTGGTTGGAACTATAACAGCCCTTAATAGAGGCTATTTCCCAACATAAAGGG	
eagand15	2059	TGATTTTCTTTTGGTTGGAACTATAACAGCCCTTAATAGAGGCTATTTCCCAACATAAAGGG	
pakd15	2075	TGATTTTCTTTTGGTTGGAACTATAACAGCCCTTAATAGAGGCTATTTCCCAACATAAAGGG	
sb33d15	2071	TGATTTTCTTTTGGTTGGAACTATAACAGCCCTTAATAGAGGCTATTTCCCAACATAAAGGG	
consensus		TGATTTTCTTTTGGTTGGAACTATAACAGCCCTTAATAGAGGCTATTTCCCAACATAAAGGG	01/02
cad15	1821	GTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTTCTGATAACAAATACTACAAAC	
minnad15	2080	GTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTTCTGATAACAAATACTACAAAC	
eagand15	2120	GTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTTCTGATAACAAATACTACAAAC	
pakd15	2136	GTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTTCTGATAACAAATACTACAAAC	
sb33d15	2132	GTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTTCTGATAACAAATACTACAAAC	
consensus		GTTAAAGCAAGTCTTGGTGGACGAGTTACTATTCCAGGTTCTGATAACAAATACTACAAAC	

FIG.1F.(CONTINUED)

cad15 1882 TAAAGTCAGATGTACAGGGTTTCTACCCATTAGACAGAGATCACCTCTGGGTTGTATCTGC
 |||||
 minnad15 2141 TAAAGTCAGATGTACAGGGTTTCTACCCATTAGACAGAGATCACCTCTGGGTTGTATCTGC
 |||||
 eagand15 2181 TAAAGTCAGATGTACAGGGTTTCTACCCATTAGACAGAGATCACCTCTGGGTTGTATCTGC
 |||||
 pakd15 2197 TAAAGTCAGATGTACAGGGTTTCTACCCATTAGACAGAGATCACCGCTGGGTTGTATCTGC
 |||||
 sb33d15 2193 TAAAGTCAGATGTACAGGGTTTCTACCCATTAGACAGAGATCACCTCTGGGTTGTATCTGC

consensus TAAAGTCAGATGTACAGGGTTTCTACCCATTAGACAGAGATCACCTCTGGGTTGTATCTGC

62/82

cad15 1943 AAAAGCATCTGCAGGATATGCAAAATGGTTTGGAAACAAGCGTTTACCGTTCTATCAAACCT
 |||||
 minnad15 2202 AAAAGCATCTGCAGGATATGCAAAATGGTTTGGAAACAAGCGTTTACCGTTCTATCAAACCT
 |||||
 eagand15 2242 AAAAGCATCTGCAGGATATGCAAAATGGTTTGGAAACAAGCGTTTACCGTTCTATCAAACCT
 |||||
 pakd15 2258 AAAAGCATCTGCAGGATATGCAAAATGGTTTGGAAACAAGCGTTTACCGTTCTATCAAACCT
 |||||
 sb33d15 2254 AAAAGCATCTGCAGGATATGCAAAATGGTTTGGAAACAAGCGTTTACCGTTCTATCAAACCT

FIG.1F.(CONTINUED)

consensus		AAAAGCATCTGCAGGATATGCAAATGGTTTGTGAAACAAGCGTTTACCGTCTCTATCAAACT
cad15	2004	TATACAGCGGTGGCATCGGTTCACTACGTGGTTTGTCTTATGGTAGTATGGACCTAACG
minnad15	2263	TATACAGCGGTGGCATCGGTTCACTACGTGGTTTGTCTTATGGTAGTATGGACCTAACG
eagand15	2303	TATACAGCGGTGGCATCGGTTCACTACGTGGTTTGTCTTATGGTAGTATGGACCTAACG
pakd15	2319	TATACAGCGGTGGCATCGGTTCACTACGTGGTTTGTCTTATGGTAGTATGGGCCCTAAAG
sb33d15	2315	TATACAGCGGTGGCATCGGTTCACTACGTGGTTTGTCTTATGGTAGCATTGGGCCCTAACG
consensus		TATACAGCGGTGGCATCGGTTCACTACGTGGTTTGTCTTATGGTAGTATTGGACCCTAACG
cad15	2065	CAATTTATGCCGAATATGGTAATGGTAGTGGTACTTTTAAGAAGATAAGTTCTGA
minnad15	2324	CAATTTATGCCGAATATGGTAATGGTAGTGGTACTTTTAAGAAGATAAGTTCTGA
eagand15	2364	CAATTTATGCCGAATATGGTAATGGTAGTGGTACTTTTAAGAAGATAAGTTCTGA
pakd15	2380	CAATTTATGCCGAACATGGTAATGGTA
		CTTTTAATAAGATAAGTTCTGA

FIG.1F.(CONTINUED)

sb33d15 2376 CAATTATcaaggtCaaaaTAAT aatTTTAATAAGATAAGTTCTGA

consensus CAATTATgccGaataTggTAATggtagtggtactggtaactTTTAAGaAGATAAGTTCTGA

cad15 2126 TGTGATTGGTGGTAATGCAATCGCTACAGCTAGCGCAGAGTTAATTGTGCCAACTCCATTT

minnad15 2385 TGTGATTGGTGGTAATGCAATCGCTACAGCTAGCGCAGAGTTAATTGTGCCAACTCCATTT

eagand15 2425 TGTGATTGGTGGTAATGCAATCGCTACAGCTAGCGCAGAGTTAATTGTGCCAACTCCATTT

pakd15 2429 TGTGATTGGTGGTAATGCAATCaCAACTGCgAGtGCAGAacTtATTGTaCCAACCTCCATTT

sb33d15 2422 TGTGATTGGTGGTAATGCAATCgCtACaGctAGcGCAGAgTtAATTGTgCCAACCTCCATTT

consensus TGTGATTGGTGGTAATGCAATCgCtACaGctAGcGCAGAgTtAATTGTgCCAACCTCCATTT

64/08

cad15 2187 GTGAGCGATAAGAGCCAAATAACGGTCCGAACCTCCTTATTGTGTGATGCGGCAAGTGTTT

minnad15 2446 GTGAGCGATAAGAGCCAAATAACGGTCCGAACCTCCTTATTGTGTGATGCGGCAAGTGTTT

eagand15 2486 GTGAGCGATAAGAGCCAAATAACGGTCCGAACCTCCTTATTGTGTGATGCGGCAAGTGTTT

consensus GTGAGCGATAAGAGCCAAATAACGGTCCGAACCTCCTTATTGTGTGATGCGGCAAGTGTTT

FIG.1F.(CONTINUED)

pkd15	2490	GTGAGTGATAaaAGCCAAAATACAGTCCGAAACCTCCCTATTTGTTGATGCGGCAAGTGTTT	
sb33d15	2483	GTGAGTGATAagAGtCAAAATACAGTCCGAAACCTCCCTATTTGTTGATGCGGCAAGTGTTT	
consensus			
		GTGAGcGATAagAGccAAAATACgGTCCGAAACCTCctTATTTGTTGATGCGGCAAGTGTTT	
cad15	2248	GGAATACTAAATGGAAATCAGATAAAAATGGATTAGAGAGCGGATGTATTAaaaAGATTGCC	
minnad15	2507	GGAATACTAAATGGAAATCAGATAAAAATGGATTAGAGAGCGGATGTATTAaaaAGATTGCC	6
			5
eagand15	2547	GGAATACTAAATGGAAATCAGATAAAAATGGATTAGAGAGCGGATGTATTAaaaAGATTGCC	8
			2
pkd15	2551	GGAATACTAAATGGAAATCAGATAAAAATGGATTAGAGAGCAagGTCCTTGAAAGACTTACC	
sb33d15	2544	GGAATACTAAATGGAAATCAGATAAAAATGGATTAGAGAGCAAtGTCCTTGAAAGACTTACC	
consensus			
		GGAATACTAAATGGAAATCAGATAAAAATGGATTAGAGAGCgAtGTaTTaaaagaTTgCC	
cad15	2309	TGATTATGGCAAAATCAAGCCGtATTCGCGCCtCTACAGGTGTCGGATTCCAAATGGCAATCT	
minnad15	2568	TGATTATGGCAAAATCAAGCCGtATTCGCGCCtCTACAGGTGTCGGATTCCAAATGGCAATCT	

FIG.1F.(CONTINUED)

eagand15 2608 TGATTATGGCAAAATCAAGCCGTATTTCGCGCCTCTACAGGTGTCGGATTCCAATGGCAATCT
 |||||
 pakd15 2612 TGATTATGGCAAAATCAAGCCGTATTTCGCGCCTCTACAGGTGTCGGATTCCAATGGCAATCT
 |||||
 sb33d15 2605 cGATTATGGCAAAATCAAGCCGTACTCGCGCCTCTACAGGTGTCGGATTCCAATGGCAATCT

consensus tGATTATGGCAAAATCAAGCCGTATTTCGCGCCTCTACAGGTGTCGGATTCCAATGGCAATCT

cad15 2370 CCTATTGGGCCATTGGTATTCTCTTATGCCAAACCAATTAAAAATATGAAAATGATGATG⁶
 |||||
 minnad15 2629 CCTATTGGGCCATTGGTATTCTCTTATGCCAAACCAATTAAAAATATGAAAATGATGATG^{6/82}
 |||||

eagand15 2669 CCTATTGGGCCATTGGTATTCTCTTATGCCAAACCAATTAAAAATATGAAAATGATGATG
 |||||
 pakd15 2673 CCTATTGGACCATTGGTATTTCTTATGCTAAACCAATTAAAAATATGAAAATGATGATG
 |||||
 sb33d15 2666 CCTAgTGGACCAGTGGTATTTCTTATGCTAAACCAATTAAAAATATGAAAATGATGATG

consensus CCTAtTGGgCCAtTGGTATTcTCTTATGcCAACCAATTAAAAATATGAAAATGATGATG

cad15 2431 TCGAACAGTTCCAAATTTAGTATTGGAGGTTCTTTCTAAATAAATTGAACTTTTTTCTTCATC
 |||||

FIG.1F.(CONTINUED)

minnad15 2690 TCGAACAGTTCCAATTAGTATTGGAGGTTCTTTCTAATAAAATTGAACTTTTTTCTTCATC
|||||
eagand15 2730 TCGAACAGTTCCAATTAGTATTGGAGGTTCTTTCTAATAAAATTGAACTTTTTTCTTCATC
|||||
pakt15 2734 TCGAACAGTTCCAATTAGTATTGGGGGCTCTTTCTAATAAAATTGAACTTTTTTCTTCATC
|||||
sb33d15 2727 TCGAACAGTTCCAATTAGTATTGGGGGCTCTTTCTAATAAAATTGAACTTTTTTCTTCATC
|||||

consensus TCGAACAGTTCCAATTAGTATTGGAGGTTCTTTCTAATAAAATTGAACTTTTTTCTTCATC

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cad15 2492 AGAACTCAAAAACAACGTTCTCTGCCCTAAATTAAATTGGGCAGAGAAAATATTAAACCCATC
|||||
minnad15 2751 AGAACTCAAAAACAACGTTCTCTGCCCTAAATTAAATTGGGCAGAGAAAATATTAAACCCATC
|||||
eagand15 2791 AGAACTCAAAAACAACGTTCTCTGCCCTAAATTAAATTGGGCAGAGAAAATATTAAACCCATC
|||||
pakt15 2795 AGAACTCAAAAACAACGTTCTCTGCCCTAAATTgAAATTGGGCAGAGAAAATATTAAACCCATC
|||||
sb33d15 2788 AGAACTCAAAAACAACGTTCTCTGCCCTAAATTtAAATTGGGCAGAGAAAATATTAAaCCATC
|||||

consensus AGAACTCAAAAACAACGTTCTCTGCCCTAAATTtAAATTGGGCAGAGAAAATATTAAACCCATC

FIG.1F.(CONTINUED)

cad15	2553	ATTTAATTAAGGATATTTATCAAAATGAAAAACATCGCAAAAGTAACCGCACCTTGCTTTTAGG	
minnad15	2812	ATTTAATTAAGGATATTTATCAAAATGAAAAACATCGCAAAAGTAACCGCACCTTGCTTTTAGG	
eagand15	2852	ATTTAATTAAGGATATTTATCAAAATGAAAAACATCGCAAAAGTAACCGCACCTTGCTTTTAGG	
pakd15	2856	ATTTAATTAAGGATATTTATCAAAATGAAAAACATCGCAAAAGTAACCGCACCTTGCTTTTAGG	
sb33d15	2849	ATTTAATTAAGGATATTTATCAAAATGAAAAACATCGCAAAAGTAACCGCACCTTGCTTTTAGG	
consensus		ATTTAATTAAGGATATTTATCAAAATGAAAAACATCGCAAAAGTAACCGCACCTTGCTTTTAGG	
600/82			
cad15	2614	TATTGCACCTTGCTTCAGGCTATGCTTCCGCTGAAGAAAAAATTGCTTTCATTAATGCAGGT	
minnad15	2873	TATTGCACCTTGCTTCAGGCTATGCTTCCGCTGAAGAAAAAATTGCTTTCATTAATGCAGGT	
eagand15	2913	TATTGCACCTTGCTTCAGGCTATGCTTCCGCTGAAGAAAAAATTGCTTTCATTAATGC ACT	
pakd15	2917	TtTTGCACCTTGCTTCAGGCTATGCTTCCGCTGAAGAAAAAATTGCTTTCATTAATGC AGG	
sb33d15	2910	TaTTGCACCTTGCTTCAGGCTATGCTgCaGCTGAAGAAAAAATTGCTTtATTAATGC AGG	
consensus		TaTTGCACCTTGCTTCAGGCTATGCTtCcGCTGAAGAAAAAATTGCTTtCATTAATGC -agt	

FIG.1F.(CONTINUED)

cad15	2675	atatttTTTcaAcatCacccagatcgccaagcggtagcagataaaacttgatgctgaatttaa	
minnad15	2934	TATAnTTTnCAAggCnaagg	
eagand15	2973	TATAtTTTCAA	
pkd15	2977	TTATATTTTtcaa	
sb33d15	2970	TTATA	
consensus		ttat-ttttcaaa-c-----gatcgccaagcggtagcagataaaacttgatgctgaatttaa	69/82
cad15	2736	acctgtagctgagaaattagcagcaagcaaaaaagaagttagataaaaattgctgctgct	
minnad15	2954		
eagand15	2985		
pkd15	2990		
sb33d15	2975		
consensus		acctgtagctgagaaattagcagcaagcaaaaaagaagttagataaaaattgctgctgct	

FIG.1F.(CONTINUED)

cad15 2797 cgtaaaaaagtagaagcaaaaagttgcggctttagaaaaagatgcacctcgcttacgtcaag

minnad15 2954

eagand15 2985

pakd15 2990

sb33d15 2975

consensus

cgtaaaaaagtagaagcaaaaagttgcggctttagaaaaagatgcacctcgcttacgtcaag

70/100

cad15 2858 ctgatatccaacacgccaacaggagattaataaattaggtgcggctgaagatgctgaatt

minnad15 2954

eagand15 2985

pakd15 2990

sb33d15 2975

consensus

ctgatatccaacacgccaacaggagattaataaattaggtgcggctgaagatgctgaatt

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FIG.1F.(CONTINUED)

cad15 2919 acaaaaattaatgcaagaacaagataaaaaa

minnad15 2954

eagand15 2985

pamd15 2990

sb33d15 2975

consensus acaaaaattaatgcaagaacaagataaaaaa

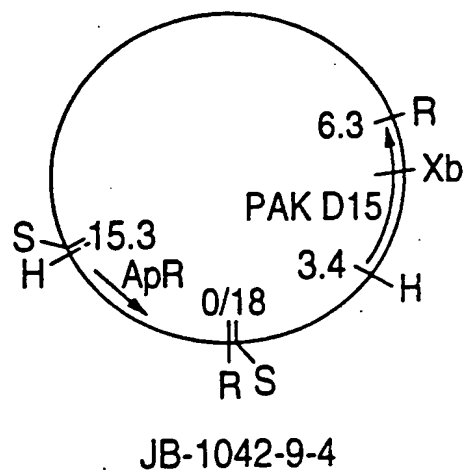
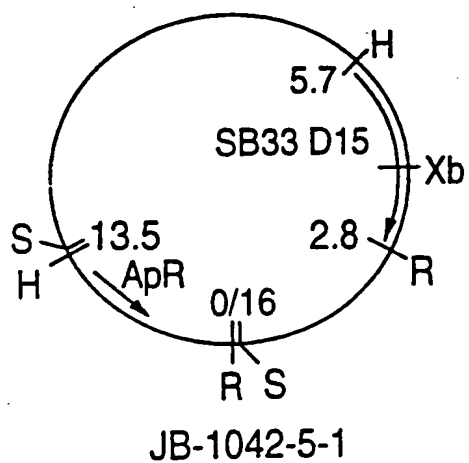
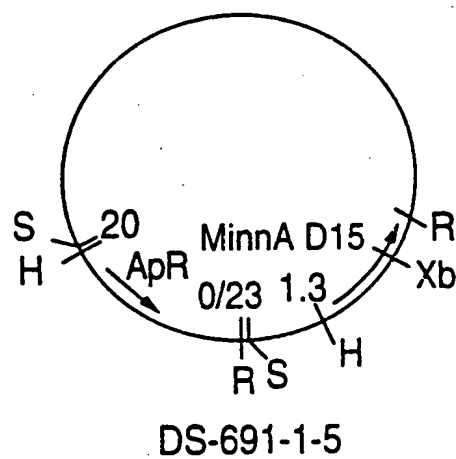
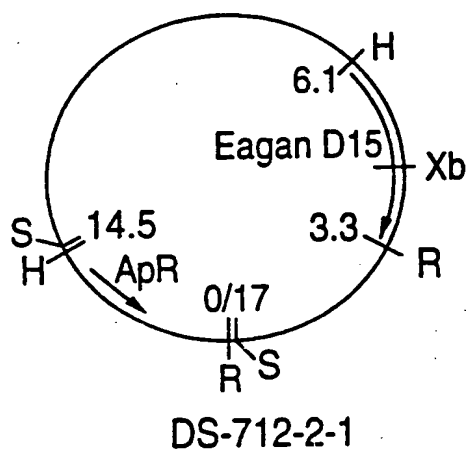
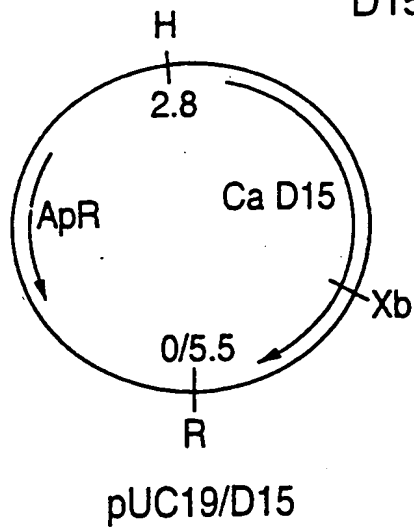
72/82
D15 CLONES

FIG.2.

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Construction of plasmid expressing SB33 D15

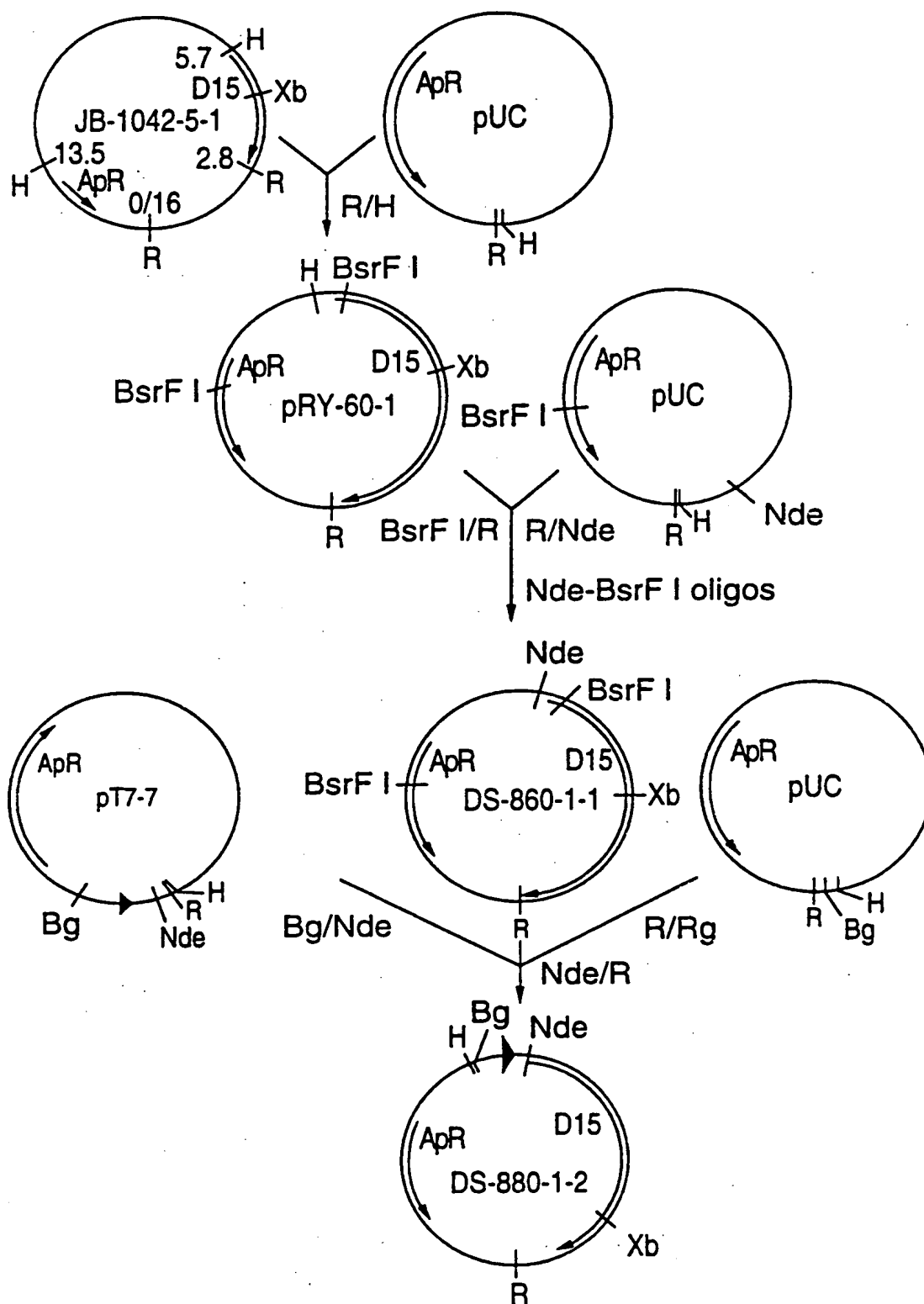
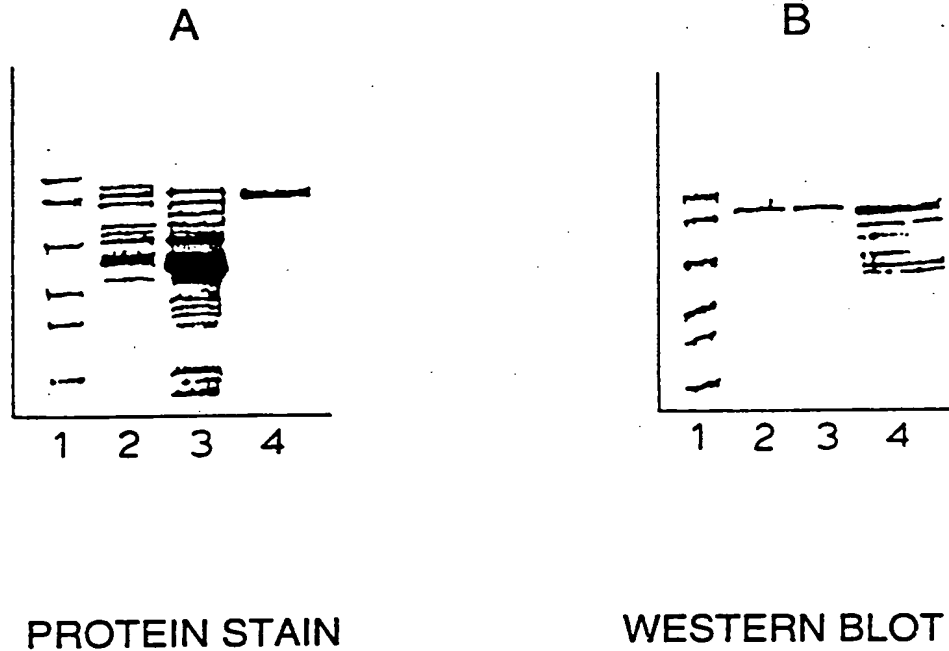


FIG.4.

76/82

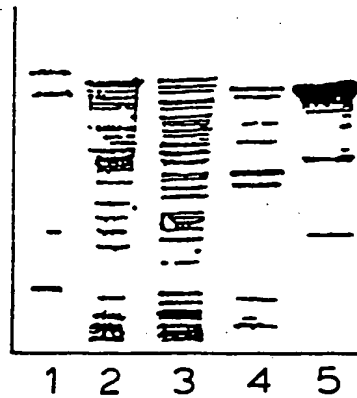
PURIFICATION OF D15 FROM A NON-TYPEABLE
HAEMOPHILUS INFLUENZAE STRAIN 30

1. Low MW markers
2. Strain 30
3. Native D15 crude extract
4. D15 after anti-D15 affinity chromatography

FIG.5.

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PURIFICATION OF FULL LENGTH RECOMBINANT D15



1. Protein M.W. Markers
2. Lysate of E. coli expressed rD15
3. Soluble protein in Tris-HCl buffer extract
4. Soluble proteins in Tris/Triton X-100/ EDTA extraction buffer
5. rD15 inclusion bodies

FIG.6.

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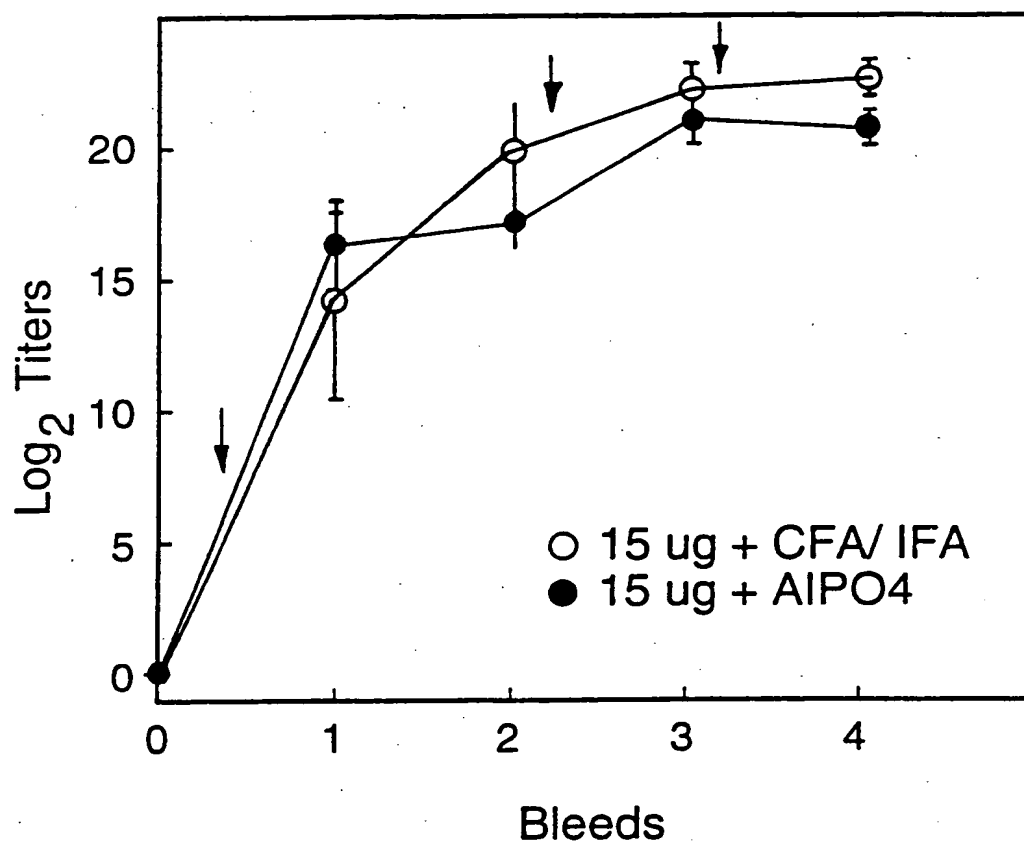


FIG.7.

79/82

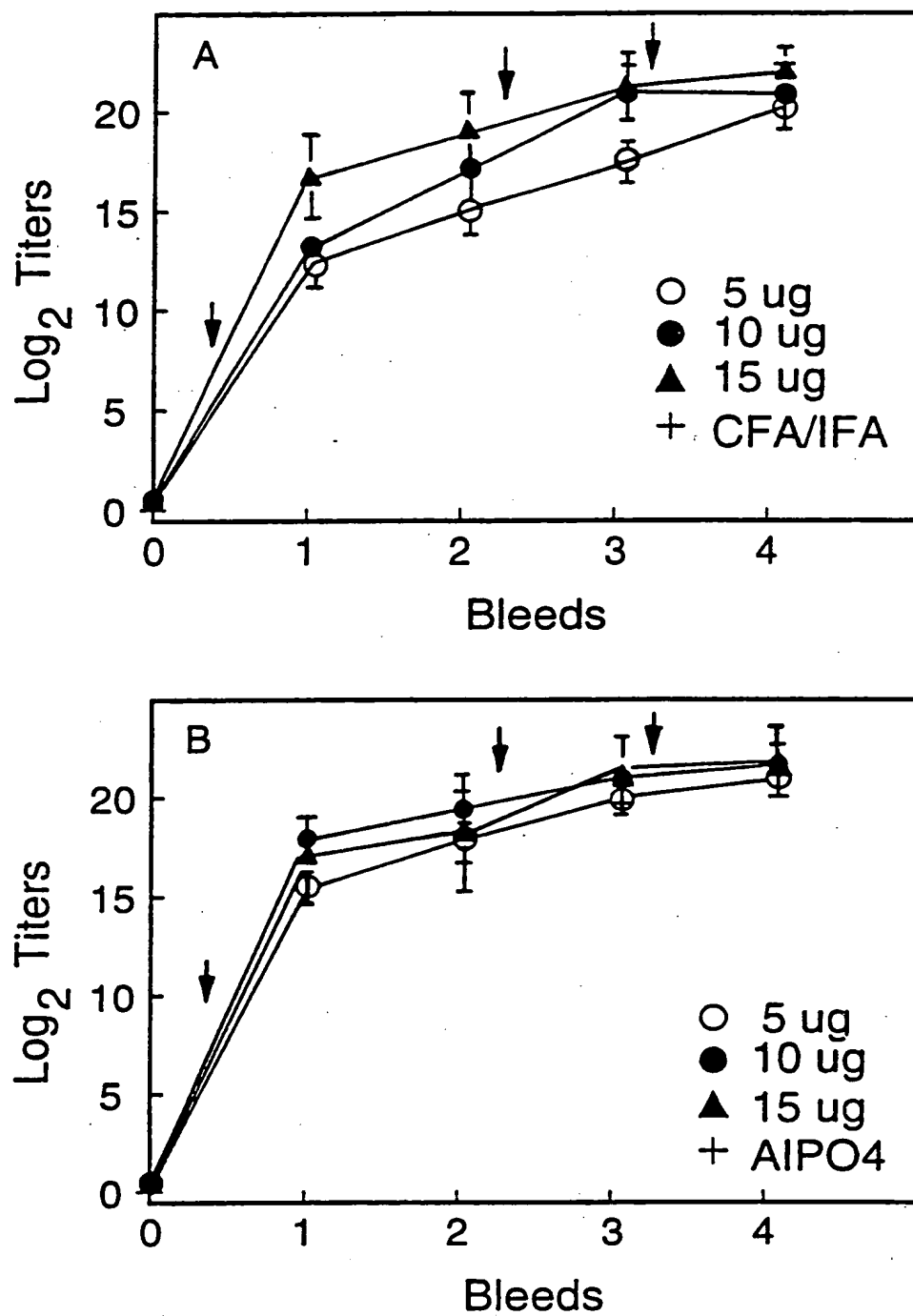
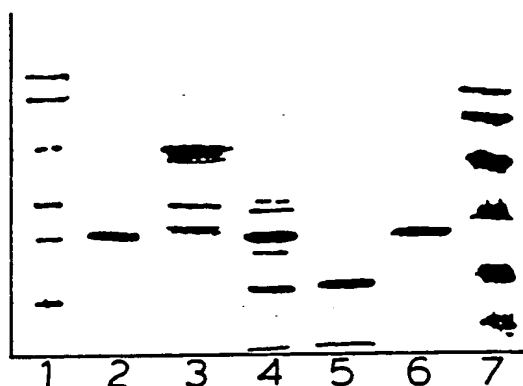


FIG.8.

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PURIFICATION OF TRUNCATED D15 FROM D15-GST
FUSION PROTEIN

1. Prestain low MW markers
2. GST standard
3. GST-(D15 fragment) fusion protein
4. Fusion protein cleaved by thrombin
5. rD15 fragment
6. GST
7. Low MW markers

FIG.9.

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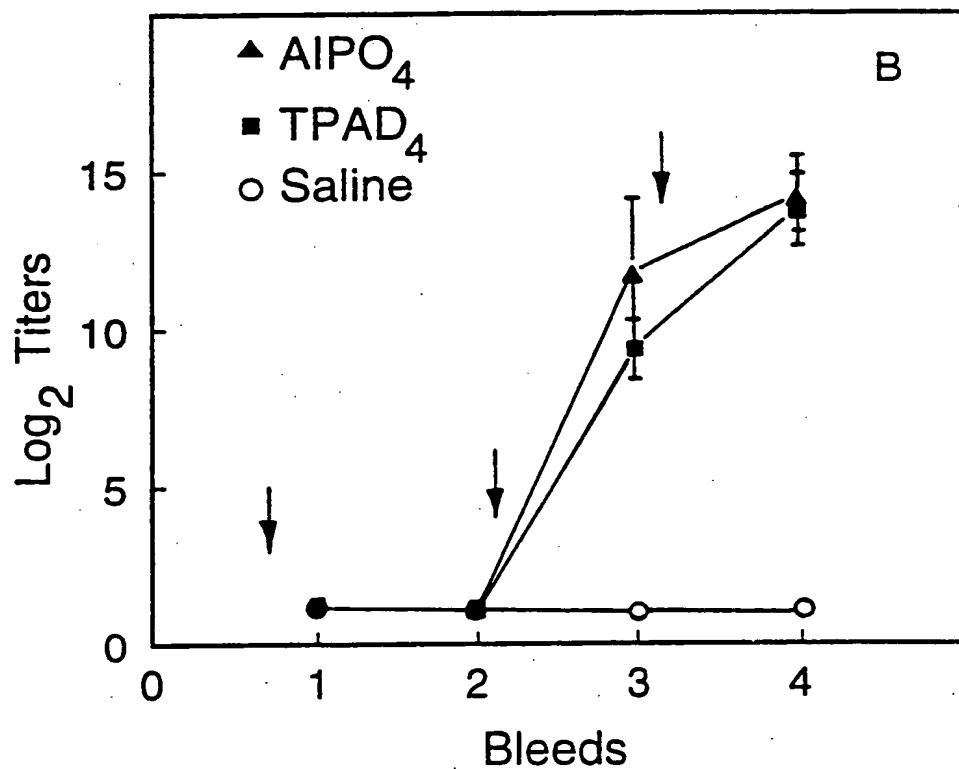
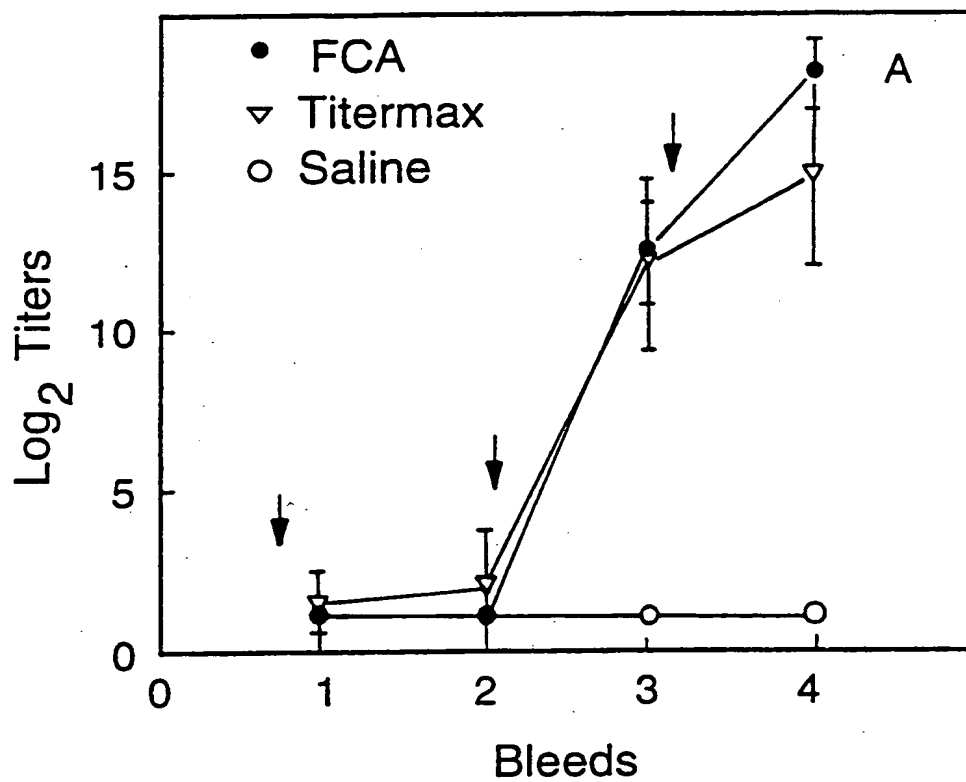


FIG.10.

82/82

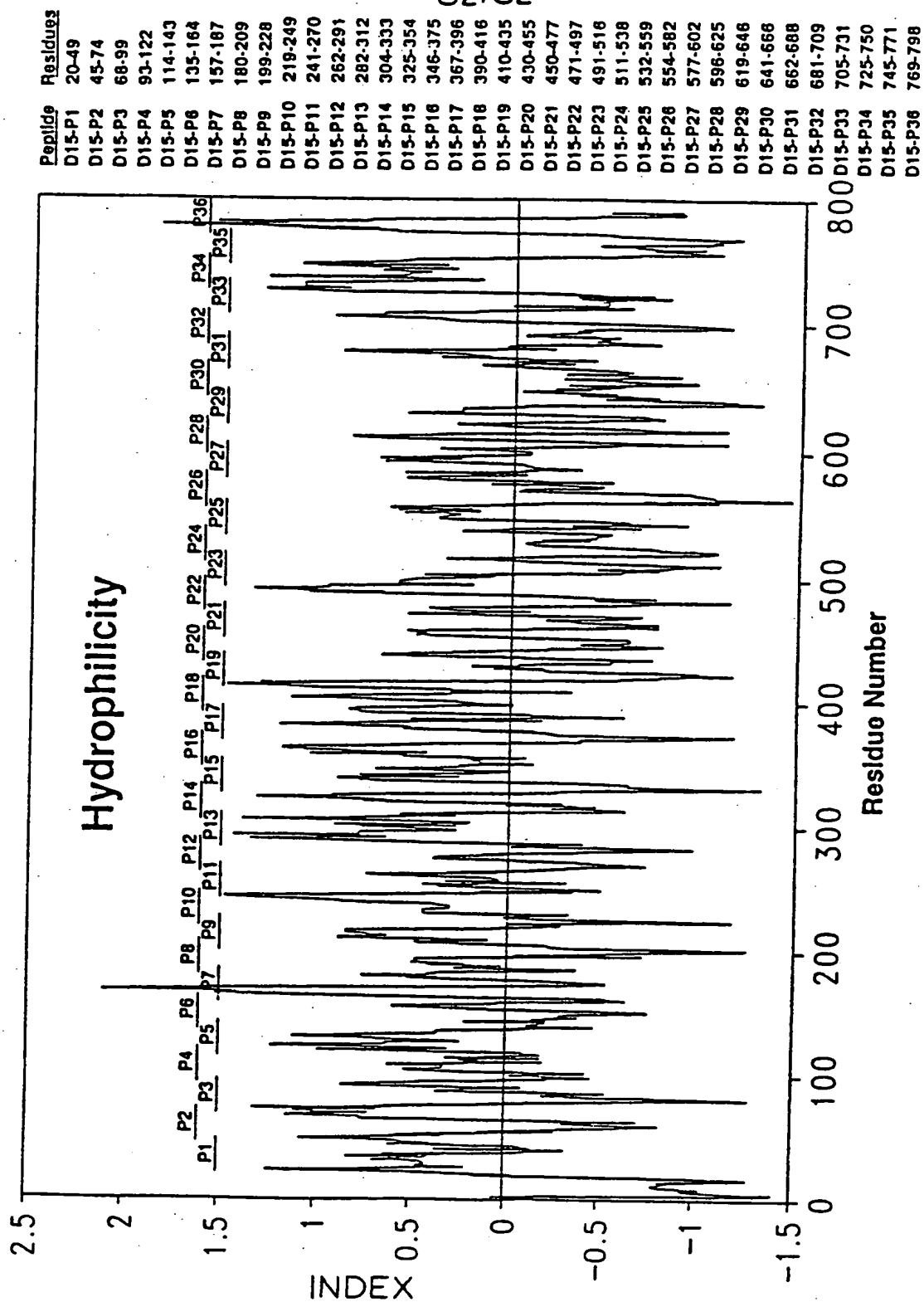


FIGURE 11

INTERNATIONAL SEARCH REPORT

International Application No

PCT/CA 93/00501

A. CLASSIFICATION OF SUBJECT MATTER

C 12 N 15/31, C 07 K 13/00, A 61 K 39/102,
 //(C 12 N 15/31; C 12 R 1/21)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

A 61 K, C 07 K, C 12 N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	INFECTION AND IMMUNITY, vol. 58, no. 4, April 1990 W.R. THOMAS et al. "Expres- sion in Escherichia coli of a High-Molecular-Weight Protective Surface Antigen Found in Nontypeable and Type b Haemophilus influ- enzae" pages 1909-1913, the whole document.	1, 4, 6, 27
A	EP, A2, 0 378 929 (CONNAUGHT LABORATORIES LIMITED) 25 July 1990 (25.07.90), claims.	1, 4, 6, 12, 16, 20, 22
A	WO, A1, 91/06 652	1, 4, 6.

☐ Further documents are listed in the continuation of box C.

☐ Patent family members are listed in annex.

* Special categories of cited documents:

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- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

15 February 1994

Date of mailing of the international search report

25 -03- 1994

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
 NL - 2280 HV Rijswijk
 Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl,
 Fax (+ 31-70) 340-3016

Authorized officer

WOLF e.h.

ANHANG

zum internationalen Recherchen-
bericht über die internationale
Patentanmeldung Nr.

ANNEX

to the International Search
Report to the International Patent
Application No.

NNEXE

au rapport de recherche inter-
national relatif à la demande de brevet
international n°

PCT/CA 93/00501 SAE 82214

In diesem Anhang sind die Mitglieder
der Patentfamilien der im obenge-
nannten internationalen Recherchenbericht
angeführten Patentdokumente angegeben.
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